



US009119851B2

(12) **United States Patent**
Bickerdike et al.

(10) **Patent No.:** **US 9,119,851 B2**
(45) **Date of Patent:** ***Sep. 1, 2015**

(54) **CYCLIC GLYCYL-2-ALLYL PROLINE
IMPROVES COGNITIVE PERFORMANCE IN
IMPAIRED ANIMALS**

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(*) Notice: Subject to any disclaimer, the term of this
patent is extended or adjusted under 35
U.S.C. 154(b) by 0 days.

This patent is subject to a terminal dis-
claimer.

(21) Appl. No.: **14/314,802**

(22) Filed: **Jun. 25, 2014**

(65) **Prior Publication Data**

US 2014/0309230 A1 Oct. 16, 2014

Related U.S. Application Data

(63) Continuation of application No. 13/043,215, filed on
Mar. 8, 2011, now Pat. No. 8,791,117, which is a
continuation-in-part of application No. 12/421,871,
filed on Apr. 10, 2009, now abandoned, which is a
continuation of application No. PCT/US2007/021744,
filed on Oct. 10, 2007, application No. 14/314,802,
which is a continuation of application No. 10/570,395,
filed as application No. PCT/US2004/028308 on Aug.
31, 2004, now Pat. No. 8,067,425.

(60) Provisional application No. 60/851,106, filed on Oct.
11, 2006, provisional application No. 60/852,507,
filed on Oct. 18, 2006, provisional application No.
60/499,956, filed on Sep. 3, 2003.

(51) **Int. Cl.**

A61K 31/498 (2006.01)

A61K 31/4985 (2006.01)

C07D 487/04 (2006.01)

C07D 487/10 (2006.01)

(52) **U.S. Cl.**

CPC **A61K 31/498** (2013.01); **A61K 31/4985**
(2013.01); **C07D 487/04** (2013.01); **C07D**
487/10 (2013.01)

(58) **Field of Classification Search**

CPC . A61K 31/401; A61K 31/407; A61K 31/498;
A61K 31/4985; C07D 403/04; C07D 487/04;
C07D 487/10

USPC 514/249; 544/349
See application file for complete search history.

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(57) **ABSTRACT**

Embodiments of this invention provide methods for therapeutic
use of cyclic G-2-Allyl Proline to treat cognitive disorders
as well as manufacture of medicaments including tablets,
capsules, injectable solutions that are useful for treatment of
such conditions.

29 Claims, 16 Drawing Sheets

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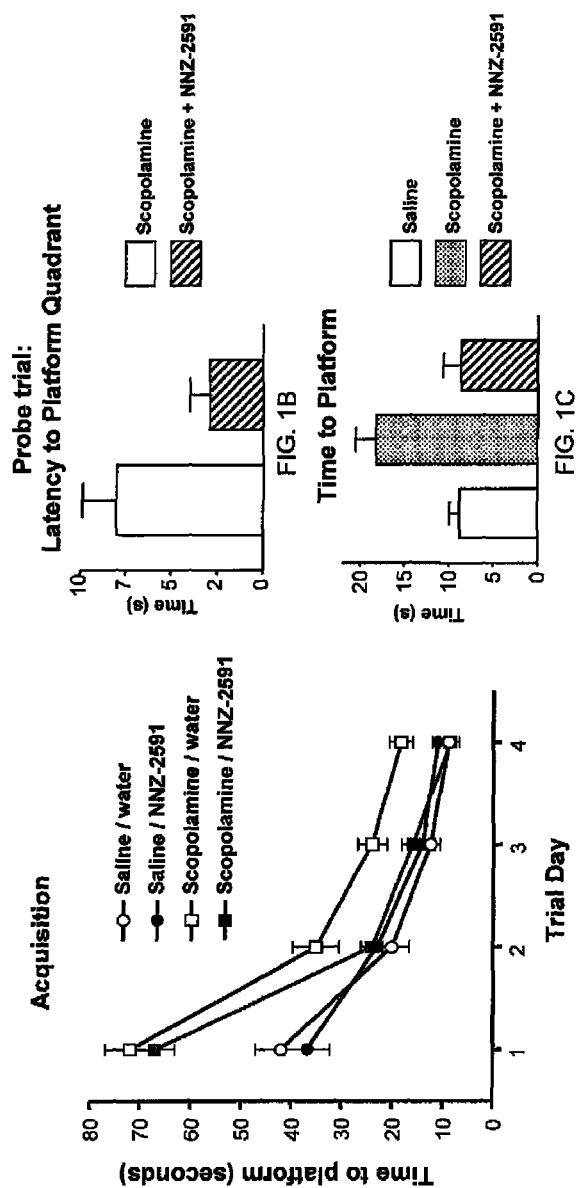


FIG. 1A

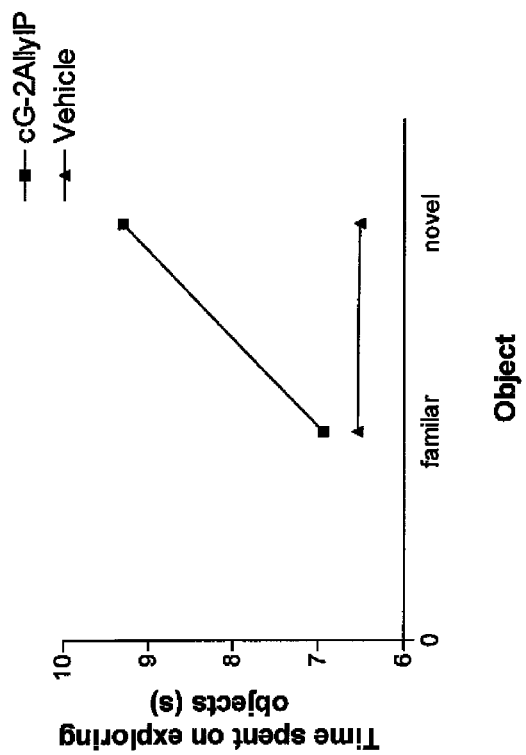


Figure 2

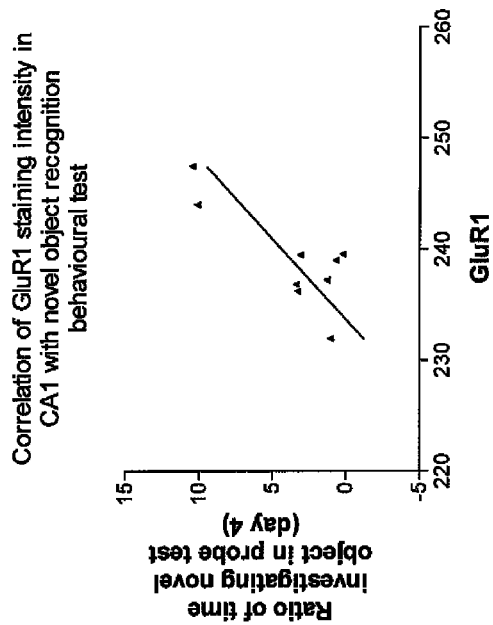


Figure 3

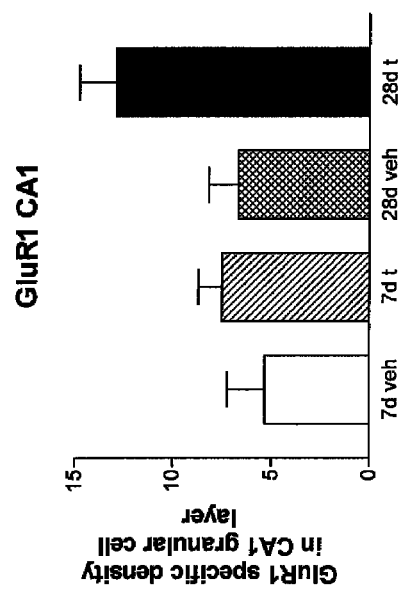


Figure 4

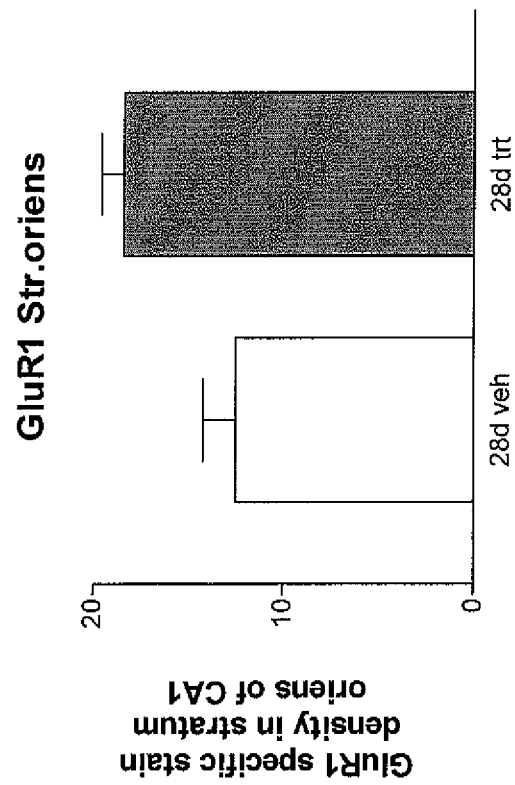


Figure 5

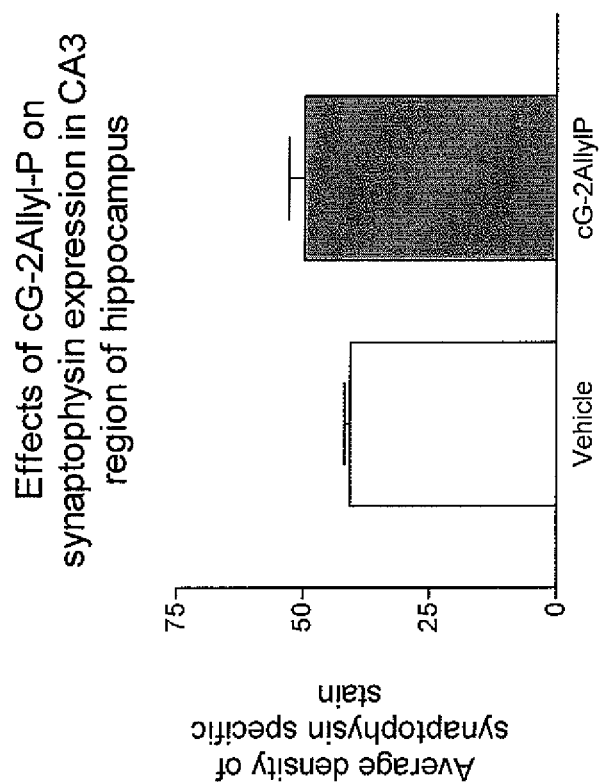


Figure 6

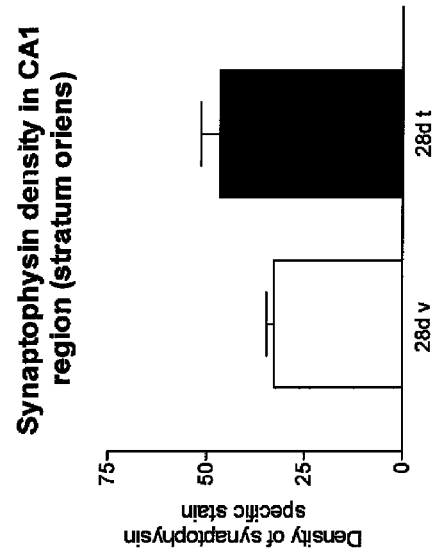


Figure 7

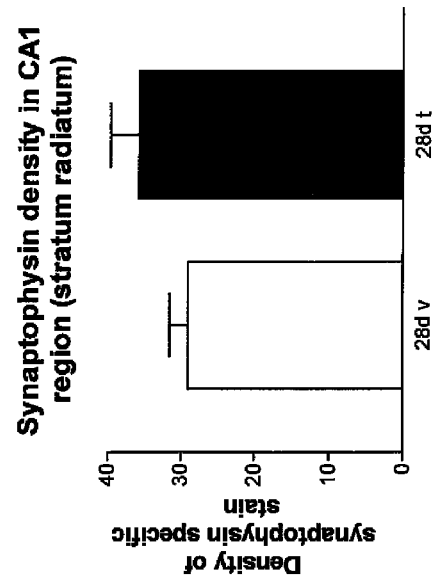


Figure 8

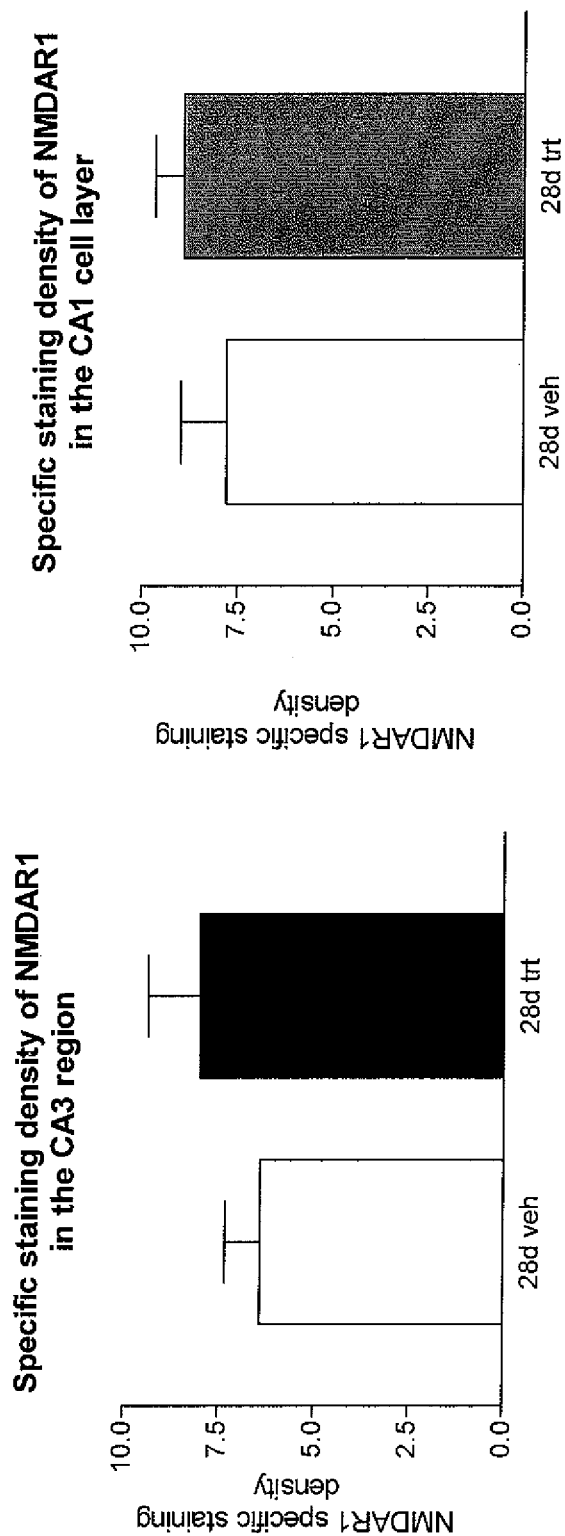


Figure 9A

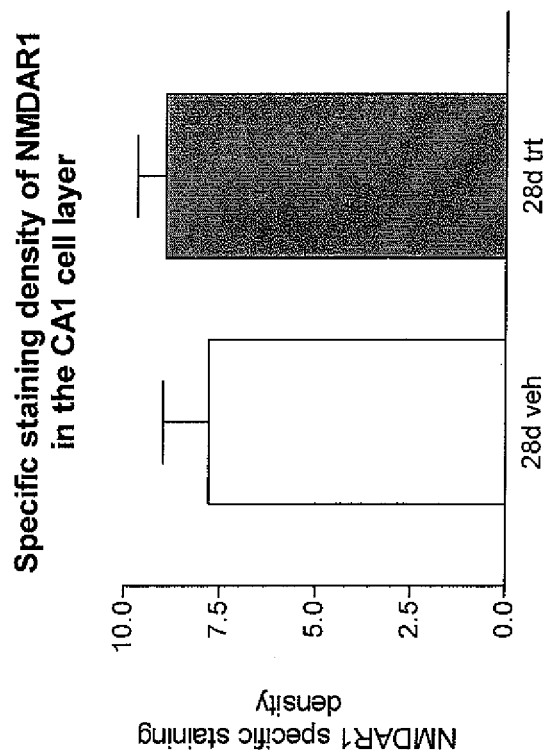


Figure 9B

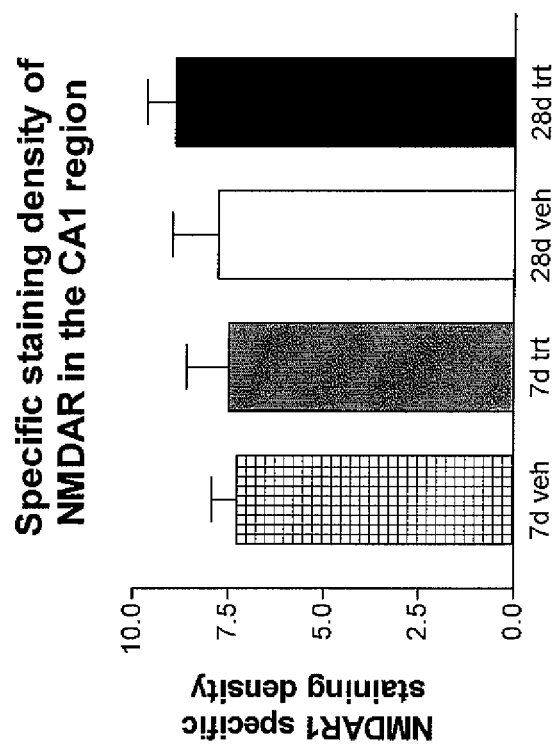


Figure 9C

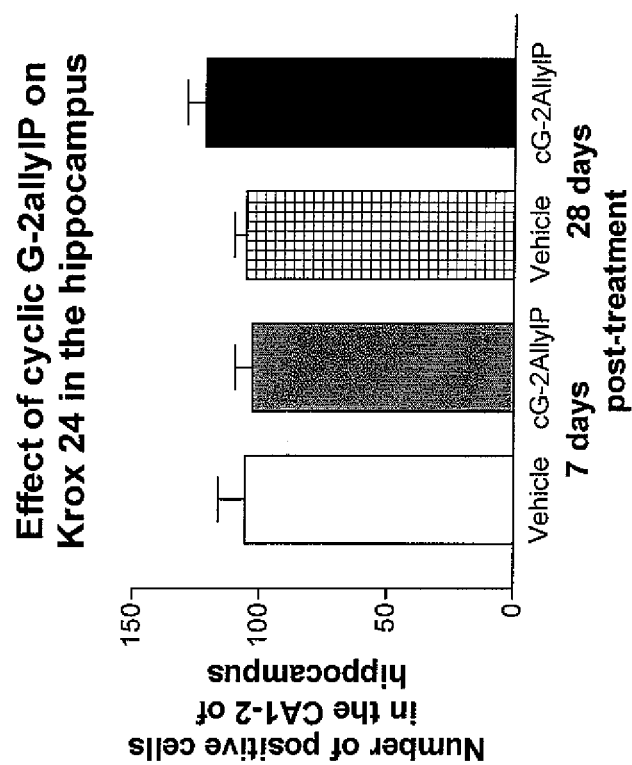


Figure 10

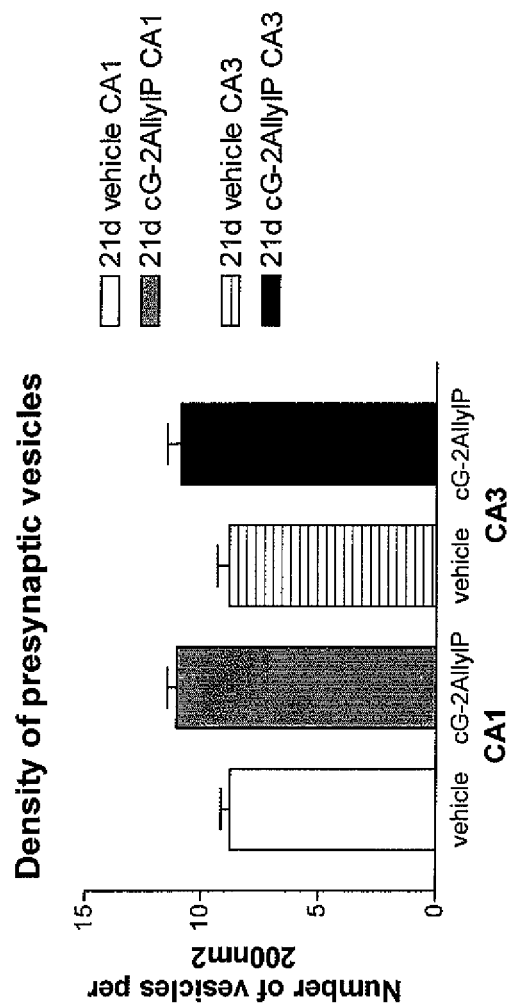
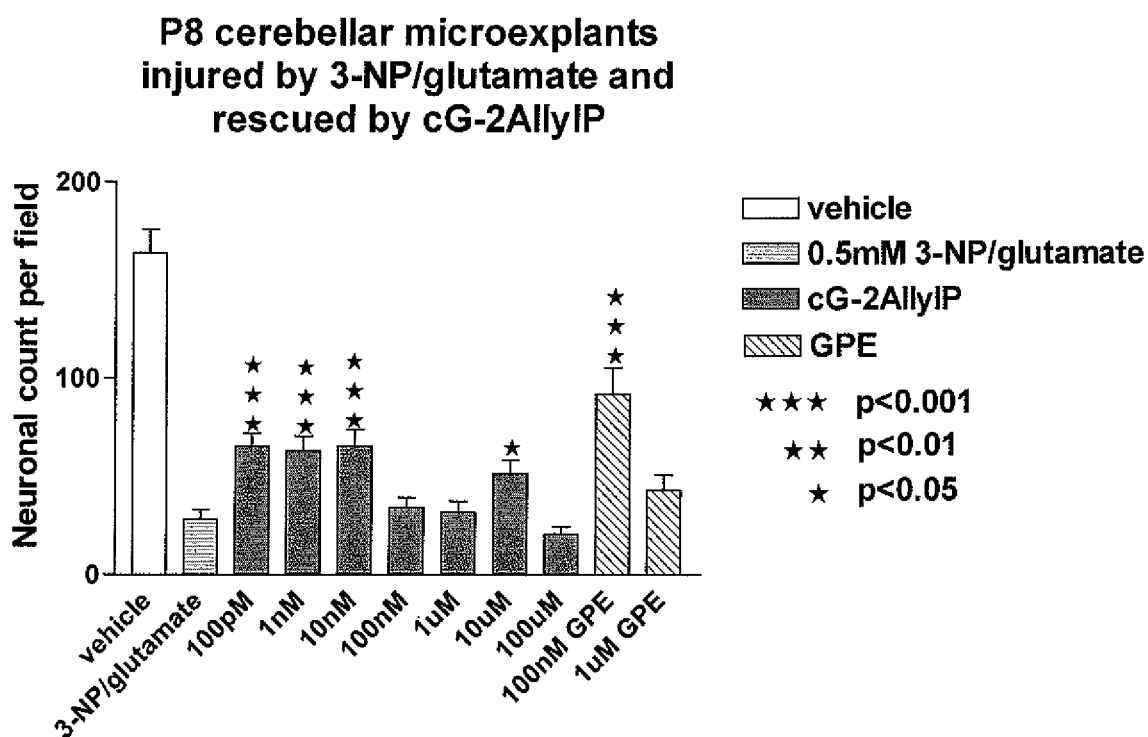
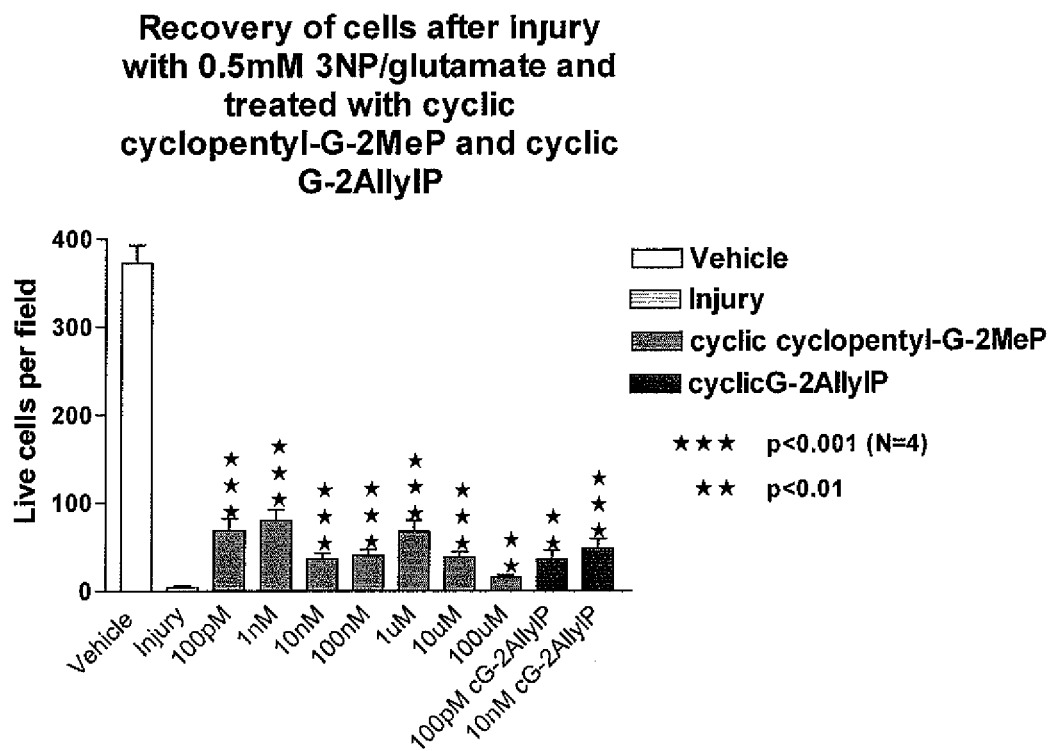


Figure 11



Cell number analysed after 48hrs.
Between 100pM to 10nM cG-2AllyIP
(N=4) the recovery from injury is in
between 25.5% to 27.3%. 100nM
GPE displays 47.1% recovery.

Figure 12



Cells are fixed after 48hrs

Figure 13

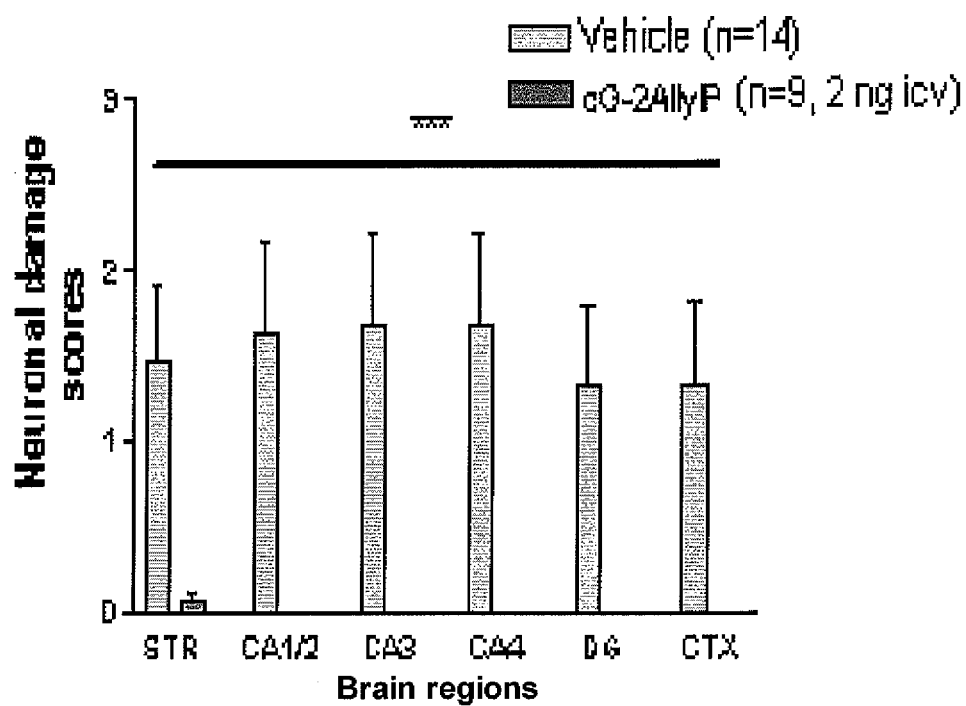
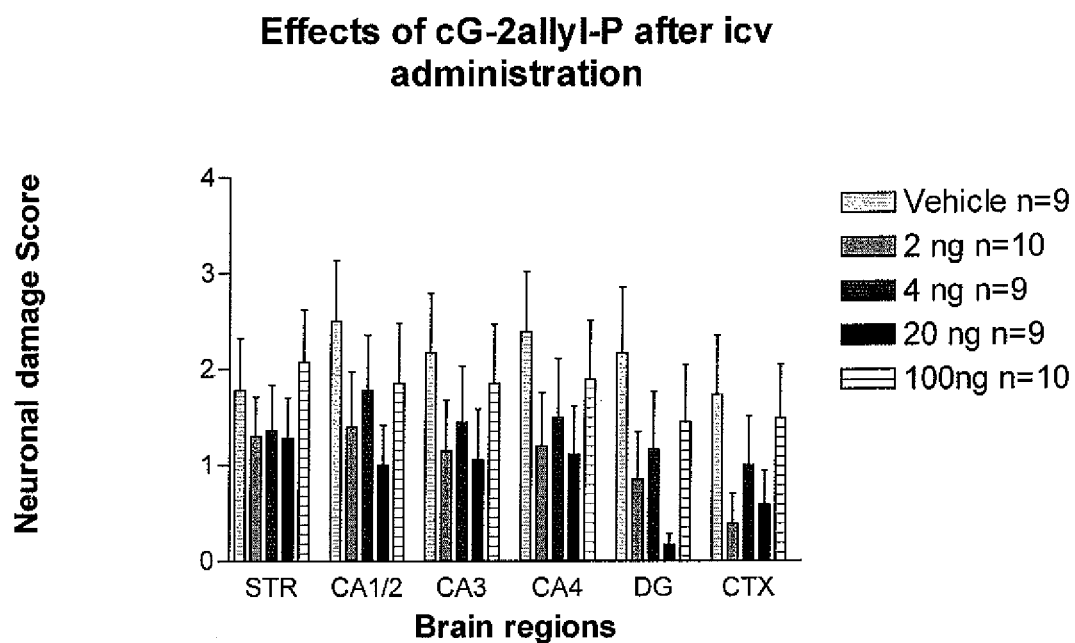


Figure 14

**Figure 15**

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CYCLIC GLYCYL-2-ALLYL PROLINE IMPROVES COGNITIVE PERFORMANCE IN IMPAIRED ANIMALS

CLAIM OF PRIORITY

This application is a Continuation of U.S. patent application Ser. No. 13/043,215, filed Mar. 8, 2011, which is a Continuation-in-Part of U.S. patent application Ser. No. 12/431,871, filed Apr. 10, 2009, now abandoned, entitled "Cyclic Glycyl-2-Allyl Proline Improves Cognitive Performance in Impaired Animals," Michael John Bickerdike and Jian Guan, inventors, which is a Continuation of PCT/US2007/021744, filed Oct. 10, 2007, now abandoned, which claims priority to U.S. Provisional Patent Application No. 60/851,106, filed Oct. 11, 2006, entitled "Cyclic Glycyl-2-Allyl Proline Improves Cognitive Performance in Impaired Animals," Michael John Bickerdike and Jian Guan, inventors, and U.S. Provisional Patent Application No. 60/852,507, filed Oct. 18, 2008, entitled "Cyclic Glycyl-2-Allyl Proline Improves Cognitive Performance in Impaired Animals," Michael John Bickerdike and Jian Guan, inventors. This application is also a Continuation-in-Part of U.S. Utility application Ser. No. 10/570,395, filed Mar. 2, 2007, now U.S. Pat. No. 8,067,425, issued 29 Nov. 2011, entitled "Neuroprotective Bicyclic Compounds and Methods for Their Use," Margaret Brimble, Jian Guan, and Frank Sieg, inventors, which is a 371(c)(1) National Phase Application of PCT/US04/28308, filed Aug. 31, 2004, now abandoned which claims priority to U.S. Provisional Patent Application No. 60/499,956, filed Sep. 3, 2003. Each of the above applications and patents are incorporated herein fully by reference.

FIELD OF THE INVENTION

The present invention relates to novel bicyclic compounds structurally related to diketopiperazines and methods for their therapeutic use. In particular, this invention relates to the neuroprotective activity of such compounds. More particularly, this invention relates to the use of cyclic Glycyl-2-Allyl Proline ("cyclic G-2-AllylP" or "cG-2-AllylP") and pharmaceutical compositions thereof in the treatment of cognitive disorders.

BACKGROUND

Cognitive disorders, i.e. impairments of memory and learning processes, have a significant detrimental effect on the quality of life of patients affected by it. Clinically recognized cognitive disorders vary from mild cognitive impairment through to dementias of varying severity.

Mild cognitive impairment ("MCI") is a transition stage between the cognitive changes of normal aging and the more serious problems caused by Alzheimer's disease. The amnesic subtype of MCI, which have been linked to development of Alzheimer's disease, significantly affects memory.

Dementia is a clinically recognised broad-spectrum syndrome consisting of degrees of loss of cognitive abilities. Dementia can be one of many symptoms of various neurological diseases or the main abnormality associated with the disease, as it is the case in Alzheimer's disease. (Adams and Victor's, Principle of Neurology, 7th ed.)

Most common causes of dementia include: cerebral atrophy associated with Alzheimer's disease, Lewy-bodies disease, frontotemporal lobar degeneration, Pick's disease; vascular narrowing or blockage in the brain (i.e. vascular

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dementia also known as multi-infarct dementia); Huntington's disease, Parkinson's disease; head trauma; HIV infection or Down's syndrome.

Currently there only several medications that have been shown to afford at most a modest transient benefit to the patients. Cholinesterase inhibitors (anticholinesterases), such as donepezil (Aricept®), galantamine (Razadyne®, Razadyne ER®, Reminyl®, Nivalin®) and rivastigmine tartrate (Exelon®) have been shown to be efficacious in mild to moderate Alzheimer's disease dementia. Exelon® has recently been approved for the treatment of mild to moderate dementia associated with Parkinson's disease. Memantine NMDA receptor antagonists are the first approved Alzheimer's disease medication acting on the glutamatergic system (Axura®, Akatinol®, Namenda®, Ebixa®). These drugs however have side effects which in some cases lead to discontinuation of the therapy.

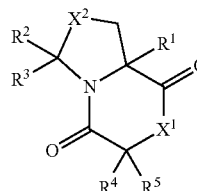
With the increase in the life span and general aging of the population there is a need to develop drugs which could delay or alleviate the cognitive function in aging patients.

SUMMARY

We have previously shown in patent application PCT/US2004/02830 filed Aug. 31, 2004, expressly incorporated herein fully by reference, that cyclic GP analogues, including but not limited to cyclic cyclopentyl-G-2-MeP and cyclic-G-2-AllylP are neuroprotective and neuroregenerative. The inventors have now discovered that cyclic G-2-AllylP is effective in treatment of cognitive impairment.

Thus, one aspect of this invention provides novel cyclic compounds having the structural formulas and substituents described below.

Formula I



In some aspects, compounds of Formula 1 include substituents where:

X¹ is selected from the group consisting of NR', O and S;
X² is selected from the group consisting of CH₂, NR', O and S;

R¹, R², R³, R⁴ and R⁵ are independently selected from the group consisting of —H, —OR', —SR', —NR'R', —NO₂, —CN, —C(O)R', —C(O)OR', —C(O)NR'R', —C(NR')NR'R', trihalomethyl, halogen, alkyl, substituted alkyl, heteroalkyl, substituted heteroalkyl, alkenyl, substituted alkenyl, alkynyl, substituted alkynyl, aryl, substituted aryl, heteroaryl, substituted heteroaryl, arylalkyl, substituted arylalkyl, heteroarylalkyl and substituted heteroarylalkyl; each R' is independently selected from the group consisting of —H, alkyl, heteroalkyl, alkenyl, alkynyl, aryl, arylalkyl, heteroaryl and heteroarylalkyl; or R⁴ and R⁵ taken together are —CH₂—(CH₂)_n—CH₂— where n is an integer from 0-6; or R² and R³ taken together are —CH₂—(CH₂)_n—CH₂— where n is an integer from 0-6; with the proviso that when R¹=methyl and R²=R³=R⁴=H then R⁵≠benzyl and; when R¹=H, at least one of R² and R³≠H.

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In further aspects, this invention provides a compound of Formula 1 or a pharmaceutically acceptable salt, stereoisomer or hydrate thereof, wherein R^1 =allyl, $R^2=R^3=R^4=R^5=H$, $X^1=NH$, $X^2=CH_2$ (cyclic Glycyl-2-AllylProline).

In still other aspects, this invention provides pharmaceutical compositions comprising a pharmaceutically acceptable excipient and a therapeutically effective amount of cyclic G-2-AllylP.

In further aspects, this invention provides methods of treating an animal having a cognitive impairment, comprising administration to that animal an effective amount of a composition comprising cyclic G-2-AllylP. In yet further aspects, the animal to be treated is a human.

BRIEF DESCRIPTION OF THE DRAWINGS

This invention is described with reference to specific embodiments thereof. Other aspects of this invention can be appreciated with reference to the drawings, in which:

FIG. 1A is a graph showing effects of treatment with cyclic G-2-AllylP on the performance in acquisition phase (days 1-4) of the Morris Water Maze Test (MWM) following scopolamine treatment. FIG. 1B is a graph showing effects of treatment with cyclic G-2-AllylP on the latency to the platform quadrant in the probe test (day 5) of the MWM. FIG. 1C is a graph showing the time taken to find the platform on day 4th of the acquisition phase for animals in 3 groups: (1) vehicle-treated, (2) scopolamine and cG-2-AllylP-treated and (3) scopolamine-treated.

FIG. 2 is a graph showing the difference in time spent on exploring the familiar vs novel object during the probe test on days 25 post-treatment. The data points for familiar objects reflect the average of time spent on exploration of 3 familiar objects. The data point for novel object recognition is the actual time spent exploring the novel object.

FIG. 3 is a graph showing a correlation between the AMPA glutamate receptor-1 staining of the CA1 region of the hippocampus and the ratio of time spent on investigation of novel object to familiar object in testing phase of the NORT on day 24.

FIG. 4 is a graph showing the effects of cG-2-AllylP (t) on the density of AMPA GluR1 in CA1 granular cell layer on days 6 and 24 in comparison to vehicle (veh).

FIG. 5 is a graph showing the effects of cG-2-AllylP (t) on the density of AMPA GluR1 in CA1 stratum oriens on day 24 post treatment.

FIG. 6 is a graph showing the effect of cG-2-AllylP on the trend to increase the density of pre-synaptic stain in CA3 region of the hippocampus at day 24 post-treatment.

FIG. 7 is a graph showing the effect of cG-2-AllylP on the trend to increase the density of the pre-synaptic stain in the stratum oriens of the CA1 region on day 24 post-treatment.

FIG. 8 is a graph showing the effect of cG-2-AllylP to increase the density of the pre-synaptic stain in the stratum radiatum of the CA1 region on day 24 post-treatment

FIGS. 9A, B, C are graphs showing the effect of cG-2-AllylP treatment on the density of the NMDAR-1 in CA1 and CA3.

FIG. 10 is a graph showing the effects of cG-2-AllylP on the density of Krox24 staining in the CA1-2 of the hippocampus.

FIG. 11 is a graph showing the effects of cG-2-AllylP on the number of vehicles in a 200 nm² square apposing the post-synaptic density in subregions CA3 and CA1 of the hippocampus of middle aged rats (n=2 in each group).

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FIG. 12 is a graph showing effects of cyclic G-2-AllylP on neuronal survival in animals following excitotoxic oxidative stress.

FIG. 13 is a graph showing effects of cyclic cyclopentylG-2-MeP on neuronal survival in animals following excitotoxic oxidative stress.

FIG. 14 is a graph showing the neuroprotective effects of cyclic G-2-AllylP in animals subjected to global brain ischaemia.

FIG. 15 is a graph showing effects of different doses of cyclic G-2-AllylP on neuroprotection in animals subjected to global brain ischaemia.

DETAILED DESCRIPTION

Definitions

“Alkenyl” refers to an unsaturated branched, straight chain or cyclic hydrocarbon radical having at least one carbon-carbon double bond. The radical may be in either the cis or trans conformation about the double bond(s). Exemplary alkenyl groups include allyl, ethenyl, propenyl, isopropenyl, butenyl, isobutenyl, cyclopentenyl and the like. In some embodiments the alkenyl groups are (C₂-C₆) alkenyl, and in other embodiments, allyl can be particularly useful.

“Alkyl” refers to a saturated branched, straight chain or cyclic hydrocarbon radical. Exemplary alkyl groups include methyl, ethyl, isopropyl, cyclopropyl, tert-butyl, cyclopropylmethyl, hexyl and the like. In some embodiments the alkyl groups are (C₁-C₆) alkyl.

“Alkynyl”, refers to an unsaturated branched, straight chain or cyclic hydrocarbon radical having at least one carbon-carbon triple bond. Exemplary alkynyl groups include ethynyl, propynyl, butynyl, isobutynyl and the like. In some embodiments the alkynyl group is (C₂-C₆) alkynyl.

“Aryl” refers to an unsaturated cyclic hydrocarbon radical with a conjugated π electron system. Exemplary aryl groups include phenyl, naphthyl and the like. In some embodiments the aryl group is (C₅-C₂₀) aryl.

“Arylalkyl” refers to a straight chain alkyl, alkenyl or alkynyl group wherein one of the hydrogen atoms bound to the terminal carbon is replaced with an aryl group. Exemplary arylalkyl groups include benzyl, naphthylmethyl, benzylidene and the like.

“Cognitive Impairment” or “Cognitive Dysfunction” means one or more signs or symptoms of memory loss, loss of spatial orientation, decreased ability to learn, decreased ability to form short- or long-term memory, decreased episodic memory, decreased ability to consolidate memory, decreased spatial memory, decreased synaptogenesis, decreased synaptic stability, deficits in cognitive mapping and scene memory, deficits in declarative and relational memory, decreased rapid acquisition of configural or conjunctive associations, decreased context-specific encoding and retrieval of specific events, decreased episodic and/or episodic-like memory. Cognitive impairment can be observed in patients having Alzheimer’s disease, Parkinson’s disease, Lewy-bodies dementia and other disorders, as well in aging animals, including humans.

“Growth factor” refers to an extracellularly active polypeptide that stimulates a cell to grow or proliferate by interacting with a receptor on the cell.

“Heteroalkyl” refers to an alkyl moiety wherein one or more carbon atoms are replaced with another atom such as N, P, O, S etc. Exemplary heteroalkyl groups include pyrrolidine, morpholine, piperidine, piperazine, imidazolidine,

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pyrazolidine, tetrahydrofuran, (C₁-C₁₀) substituted amines, (C₂-C₆) thioethers and the like.

"Heteroaryl" refers to an aryl moiety wherein one or more carbon atoms are replaced with another atom such as N, P, O, S etc. Exemplary heteroaryl groups include carbazole, furan, imidazole, indazole, indole, isoquinoline, purine, pyrazine, pyrazole, pyridazine, pyridine, pyrrole, thiazole, thiophene, triazole and the like.

"Injury" includes any acute or chronic damage of an animal that results in degeneration or death of cells in the nervous system. Such cells include neuronal cells and non-neuronal cells. Injury includes stroke, non-hemorrhagic stroke, traumatic brain injury, perinatal asphyxia associated with fetal distress such as following abruption, cord occlusion or associated with intrauterine growth retardation, perinatal asphyxia associated with failure of adequate resuscitation or respiration, severe CNS insults associated with near miss drowning, near miss cot death, carbon monoxide inhalation, ammonia or other gaseous intoxication, cardiac arrest, coma, meningitis, hypoglycemia and status epilepticus, episodes of cerebral asphyxia associated with coronary bypass surgery, hypotensive episodes and hypertensive crises, and cerebral trauma. It is to be understood that the above examples are by way of illustration only, and are not intended to be a complete listing of injuries capable of being treated by the compounds and methods of this invention.

A "pharmaceutically acceptable excipient" refers to an excipient that is useful in preparing a pharmaceutical composition that is generally safe, non-toxic, and desirable, and includes excipients that are acceptable for veterinary use as well as for human pharmaceutical use. Such excipients may be solid, liquid, semisolid, or, in the case of an aerosol composition, gaseous.

A "pharmaceutically acceptable salt" refers to a salt that is pharmaceutically acceptable and has the desired pharmacological properties. Such salts include salts that may be formed where acidic protons present in the compounds are capable of reacting with inorganic or organic bases. Suitable inorganic salts include those formed with the alkali metals, e.g. sodium and potassium, magnesium, calcium, and aluminium. Suitable organic salts include those formed with organic bases such as the amine bases e.g. ethanolamine, diethanolamine, triethanolamine, tromethamine, N-methylglucamine, and the like. Such salts also include acid addition salts formed with inorganic acids (e.g. hydrochloric and hydrobromic acids) and organic acids (e.g. acetic acid, citric acid, maleic acid, and the alkane- and arene-sulfonic acids such as methanesulfonic acid and benzenesulfonic acid). When there are two acidic groups present, a pharmaceutically acceptable salt may be a mono-acid mono-salt or a di-acid salt; and similarly where there are more than two acidic groups present, some or all of such groups can be present as salts.

A "protecting group" has the meaning conventionally associated with it in organic synthesis, i.e. a group that selectively blocks one or more reactive sites in a multifunctional compound such that a chemical reaction can be carried out selectively on another unprotected reactive site and such that the group can readily be removed after the selective reaction is complete.

A "stereoisomer" is a molecule having the structure of cyclic G-2-Allyl Proline, but having a chiral center. The term "cyclic G-2-Allyl Proline" includes all stereoisomers.

"Substituted" refers to where one or more of the hydrogen atoms on an alkyl, heteroalkyl, alkenyl, alkynyl, aryl, heteroaryl or arylalkyl radical are independently replaced with another substituent. Substituents include —R', —OR', —SR', —NR'R', —NO₂, —CN, —C(O)R', —C(O)OR', —C(O)

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NR'R', —C(NR')NR'R', —NR'—C(NR)—OR', —NR'—C(NR)—SR', NR'—C(NR')—NR'R', trihalomethyl and halogen where each R' is independently —H, alkyl, heteroalkyl, alkenyl, alkynyl, aryl, arylalkyl, heteroaryl or heteroarylalkyl.

A "therapeutically effective amount" means the amount that, when administered to an animal for treating a disease, is sufficient to effect treatment for a disease or an injury. A "therapeutically effective amount" means an amount that decreases adverse symptoms or findings, promotes desirable symptoms or findings, and/or treats an underlying disorder, and/or is curative.

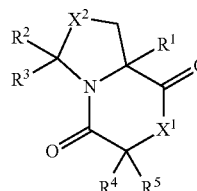
"Treating" or "treatment" of a disease includes preventing the disease from occurring in an animal that may be predisposed to the disease but does not yet experience or exhibit symptoms of the disease (prophylactic treatment), inhibiting the disease (slowing or arresting its development), providing relief from the symptoms or side-effects of the disease (including palliative treatment), and relieving the disease (causing regression of the disease).

Implicit hydrogen atoms (such as the hydrogens on the pyrrole ring, etc.) are omitted from the formulae for clarity, but should be understood to be present.

Compounds of the Invention

Certain embodiments of this invention include novel derivatives of cPG having structures as described below.

Formula I



In certain embodiments, compounds of Formula 1 include substituents where:

X¹ is selected from the group consisting of NR', O and S;

X² is selected from the group consisting of CH₂, NR', O and S;

R¹, R², R³, R⁴ and R⁵ are independently selected from the group consisting of —H, —OR', —SR', —NR'R', —NO₂, —CN, —C(O)R', —C(O)OR', —C(O)NR'R', —C(NR')NR'R', trihalomethyl, halogen, alkyl, substituted alkyl, heteroalkyl, substituted heteroalkyl, alkenyl, substituted alkenyl, alkynyl, substituted alkynyl, aryl, substituted aryl, heteroaryl, substituted heteroaryl, arylalkyl, substituted arylalkyl, heteroarylalkyl and substituted heteroarylalkyl; each R' is independently selected from the group consisting of —H, alkyl, heteroalkyl, alkenyl, alkynyl, aryl, arylalkyl, heteroaryl and heteroarylalkyl;

or R⁴ and R⁵ taken together are —CH₂—(CH₂)_n—CH₂— where n is an integer from 0-6;

or R² and R³ taken together are —CH₂—(CH₂)_n—CH₂— where n is an integer from 0-6; with the proviso that when R¹=methyl and R²=R³=R⁴=H then R⁵≠benzyl and; when R¹=H, at least one of R² and R³≠H.

In further embodiments, compounds of Formula 1 include substituents where:

R¹=methyl, R²=R³=R⁴=R⁵=H, X²=CH₂;

R¹=allyl, R²=R³=R⁴=R⁵=H, X¹=NH, X²=CH₂;

R¹=R²=R³=H, R⁴=R⁵=methyl, X¹=NH, X²=CH₂;

R¹=R²=R³=methyl, X¹=NH, X²=CH₂.

In other embodiments of the invention, compounds of Formula 1 include substituents where;

R^4 and R^5 taken together are $-\text{CH}_2-(\text{CH}_2)_n-\text{CH}_2-$ and:

$R^1=\text{methyl}$, $R^2=R^3=\text{H}$, $n=0$, $X^1=\text{NH}$, $X^2=\text{CH}_2$;

$R^1=\text{methyl}$, $R^2=R^3=\text{H}$, $n=2$, $X^1=\text{NH}$, $X^2=\text{CH}_2$;

$R^1=\text{allyl}$, $R^2=R^3=\text{H}$, $n=0$, $X^1=\text{NH}$, $X^2=\text{CH}_2$;

$R^1=\text{allyl}$, $R^2=R^3=\text{H}$, $n=2$, $X^1=\text{NH}$, $X^2=\text{CH}_2$;

$R^1=\text{methyl}$, $R^2=R^3=\text{H}$, $n=3$, $X^1=\text{NH}$, $X^2=\text{CH}_2$;

$R^1=\text{allyl}$, $R^2=R^3=\text{H}$, $n=3$, $X^1=\text{NH}$, $X^2=\text{CH}_2$.

In still other embodiments of the invention, compounds of Formula 1 include substituents where $R^1=\text{methyl}$ or allyl , $R^2=R^3=R^4=\text{H}$ and R^5 is selected from the group consisting of the side chains of the amino acids: alanine, arginine, asparagine, aspartic acid, cysteine, glutamic acid, glutamine, histidine, isoleucine, leucine, lysine, methionine, proline, serine, threonine, tryptophan, tyrosine, valine, norvaline, norleucine, citrulline, ornithine, homocysteine, homoserine, alloisoleucine, isovaline, sarcosine and the like.

In yet further embodiments of the invention, compounds of Formula 1 include substituents where:

$R^1=\text{methyl}$, $R^2=R^3=\text{methyl}$, $R^4=R^5=\text{H}$, $X^1=\text{NH}$ and $X^2=\text{S}$;

$R^1=\text{allyl}$, $R^2=R^3=\text{methyl}$, $R^4=R^5=\text{H}$, $X^1=\text{NH}$ and $X^2=\text{S}$.

Those with skill in the art will appreciate that the above structural representations can contain chiral centres, the number of which will depend on the different substituents. The chirality may be either R or S at each centre. The structural drawings can represent only one of the possible tautomeric, conformational diastereomeric or enantiomeric forms, and it should be understood that the invention encompasses any tautomeric, conformational isomeric diastereomeric or enantiomeric form, which exhibits biological or pharmacological activity as described herein.

Pharmacology and Utility

Cyclic Glycyl-2-Allyl proline (cG-2-AllylP) is described in U.S. Utility application Ser. No. 11/399,974 filed Apr. 7, 2006, entitled "Cyclic G-2-Allyl Proline in Treatment of Parkinson's Disease," now U.S. Pat. No. 7,776,876, issued Aug. 17, 2010, U.S. Utility application Ser. No. 10/570,395, filed Mar. 2, 2006 entitled "Neuroprotective Bicyclic Compounds and Methods for Their Use", PCT International Patent Application No: PCT/US2004/028308, entitled Neuroprotective Bicyclic Compounds and methods for Their Use, and U.S. Provisional Patent Application Ser. No. 60/499,956 filed Sep. 3, 2003, entitled "Neuroprotective Bicyclic Compounds and Methods for Their Use". Each of the above patent applications and the patent is expressly incorporated herein fully by reference.

Certain aspects of this invention include the use of cyclic G-2-AllylP in treatment of cognitive impairment associated with aging with neurodegenerative conditions or in situations in which cognitive impairment is found with no apparent neurodegeneration.

Scopolamine is commonly used in animal models of cholinergic hypofunction associated with Alzheimer's disease. The functional deficits observed after scopolamine treatment include those found in human patients with Alzheimer's disease. Thus, scopolamine treatment is reasonably predictive of cognitive impairment found in human diseases. Additionally, scopolamine treatment mimics cognitive dysfunction in humans who do not have neurodegenerative disorders.

cG-2-AllylP administered to animals treated with scopolamine-induced cognitive dysfunction produces clinical improvement in those animals, similar to the therapeutic improvement observed in people suffering from cholinergic hypofunction. For example, cholinergic hypofunction associated with Alzheimer's disease. Thus, studies of effects of

Cyclic G-2-AllylP scopolamine treated animals are reasonably predictive of effects observed in human beings suffering from cholinergic dysfunction.

Such other agents may be selected from the group consisting of for example, growth factors and associated derivatives, e.g., insulin-like growth factor-I (IGF-I), insulin-like growth factor-II (IGF-II), the tripeptide GPE, transforming growth factor- β 1, activin, growth hormone, nerve growth factor, growth hormone binding protein, and/or IGF-binding proteins. Additional compounds include Glycyl-2-Methyl Prolyl Glutamate and/or other compounds disclosed in U.S. patent application Ser. No. 10/155,864, now U.S. Pat. No. 7,041,314, issued May 9, 2006, expressly incorporated herein fully by reference.

Therapeutic Applications

Compositions and methods of the invention find use in the treatment of animals, such as human patients, suffering from cognitive impairment. Still more generally, the compositions and methods of the invention find use in the treatment of mammals, such as human patients, suffering from memory impairment, mild cognitive impairment, dementia, including dementia including dementias resulting from cerebral atrophy associated with Alzheimer's disease, Lewy-bodies disease, frontotemporal lobar degeneration, Pick's disease; vascular narrowing or blockage in the brain (i.e. vascular dementia also known as multi-infarct dementia); Huntington's disease, Parkinson's disease; head trauma; HIV infection or Down's syndrome.

Pharmaceutical Compositions and Administration

Cyclic G-2-AllylP can be administered as part of a medicament or pharmaceutical preparation. This can involve combining a compound of the invention with any pharmaceutically appropriate carrier, adjuvant or excipient. The selection of the carrier, adjuvant or excipient will of course usually be dependent upon the route of administration to be employed.

In general, compounds of this invention will be administered in therapeutically effective amounts by any of the usual modes known in the art, either singly or in combination with other conventional therapeutic agents for the disease being treated. A therapeutically effective amount may vary widely depending on the disease or injury, its severity, the age and relative health of the animal being treated, the potency of the compound(s), and other factors. As anti-apoptotic and anti-necrotic agents, therapeutically effective amounts of cyclic G-2-AllylP may range from 0.001 to 100 milligrams per kilogram mass of the animal, with lower doses such as 0.001 to 0.1 mg/kg being appropriate for administration through the cerebrospinal fluid, such as by intracerebroventricular administration, and higher doses such as 1 to 100 mg/kg being appropriate for administration by methods such as oral, systemic (e.g. transdermal), or parenteral (e.g. intravenous) administration. A person of ordinary skill in the art will be able without undue experimentation, having regard to that skill and this disclosure, to determine a therapeutically effective amount of a compound of this invention for a given disease or injury.

Cyclic G-2-AllylP may be administered peripherally via any peripheral route known in the art. These can include parenteral routes for example injection into the peripheral circulation, subcutaneous, intraorbital, ophthalmic, intraspinal, intracisternal, topical, infusion (using e.g. slow release devices or minipumps such as osmotic pumps or skin patches), implant, aerosol, inhalation, scarification, intraperitoneal, intracapsular, intramuscular, intranasal, oral, buccal, transdermal, pulmonary, rectal or vaginal. The compositions can be formulated for parenteral administration to humans or other mammals in therapeutically effective amounts (e.g.

amounts which eliminate or reduce the patient's pathological condition) to provide therapy for the neurological diseases described above.

Desirably, if possible, when administered as anti-apoptotic and anti-necrotic agent, cyclic G-2-AllylP can be administered orally. The amount of a compound of this invention in the composition may vary widely depending on the type of composition, size of a unit dosage, kind of excipients, and other factors well known to those of ordinary skill in the art. In general, the final composition may comprise from 0.0001 percent by weight (% w) to 10% w of the compound of this invention, preferably 0.001% w to 1% w, with the remainder being the excipient or excipients.

Other convenient administration routes include subcutaneous injection (e.g. dissolved in a physiologically compatible carrier such as 0.9% sodium chloride) or direct administration to the CNS. Using stereotactic devices and accurate maps of an animals' CNS, a compound may be injected directly into a site of neural damage. Such routes of administration may be especially desired in situations in which perfusion of that location is compromised either by decreased vascular perfusion or by decreased cerebral spinal fluid (CSF) flow to that area. Examples include administration by lateral cerebroventricular injection or through a surgically inserted shunt into the lateral cerebroventricle of the brain of the patient, intravenously, direct injection into the desired location or other routes.

The effective amount of compound in the CNS may be increased by administration of a pro-drug form of a compound, which comprises a compound of the invention and a carrier, where the carrier is joined to a compound of the invention by a linkage which is susceptible to cleavage or digestion within the patient. Any suitable linkage can be employed which will be cleaved or digested following administration.

However, there is no intention on the part of the applicants to exclude other forms of administration.

In further embodiments of the invention, restoring nerve function in an animal can comprise administering a therapeutic amount of cyclic G-2-AllylP in combination with another neuroprotective agent, selected from, for example, growth factors and associated derivatives (insulin-like growth factor-I (IGF-I), insulin-like growth factor-II (IGF-II), transforming growth factor- β 1, activin, growth hormone, nerve growth factor, growth hormone binding protein, IGF-binding proteins (especially IGFBP-3), basic fibroblast growth factor, acidic fibroblast growth factor, the hst/Kfgk gene product, FGF-3, FGF-4, FGF-6, keratinocyte growth factor, androgen-induced growth factor. Additional members of the FGF family include, for example, int-2, fibroblast growth factor homologous factor-1 (FHF-1), FHF-2, FHF-3 and FHF-4, keratinocyte growth factor 2, glial-activating factor, FGF-10 and FGF-16, ciliary neurotrophic factor, brain derived growth factor, neurotrophin 3, neurotrophin 4, bone morphogenetic protein 2 (BMP-2), glial-cell line derived neurotrophic factor, activity-dependant neurotrophic factor, cytokine leukaemia inhibiting factor, oncostatin M, interleukin), α -, β -, γ -, or consensus interferon, and TNF- α . Other forms of neuroprotective therapeutic agents include, for example, clomethiazole; kynurenic acid, Semax, tacrolimus, L-threo-1-phenyl-2-decanoylamino-3-morpholino-1-propanol, and adrenocorticotropin-(4-9) analogue (ORG 2766) and dizolciline (MK-801), selegiline; glutamate antagonists such as, NPS1506, GV1505260, MK-801, GV150526; AMPA antagonists such as 2,3-dihydroxy-6-nitro-7-sulfamoylbenzo (f)quinoxaline (NBQX), LY303070 and LY300164; anti-inflammatory agents directed against the addressin MAd-

CAM-1 and/or its integrin α 4 receptors (α 4 β 1 and α 4 β 7), such as anti-MAdCAM-1 mAb MECA-367 (ATCC accession no. HB-9478).

Cyclic G-2-AllylP is suitably administered by a sustained-release system. Suitable examples of sustained-release compositions include semi-permeable polymer matrices in the form of shaped articles, e.g., films, or microcapsules. Sustained-release matrices include polylactides (U.S. Pat. No. 3,773,919; EP 58,481), copolymers of L-glutamic acid and gamma-ethyl-L-glutamate (Sidman et al., 1983, *Biopolymers*: 22: 547-56), poly(2-hydroxyethyl methacrylate) (Langer et al., 1981, *J. Biomed. Mater. Res.*: 15: 267), ethylene vinyl acetate (Langer et al., 1981, *J. Biomed. Mater. Res.*: 15: 267), or poly-D-(-)-3-hydroxybutyric acid (EP 133,988). Sustained-release compositions also include a liposomally entrapped compound. Liposomes containing the compound are prepared by methods known per se: DE 3,218,121, EP 52,322, EP 36,676, EP 88,046, EP 143,949, EP 142,641, Japanese Pat. Appln. 83-118008, U.S. Pat. Nos. 4,485,045 and 4,544,545, and EP 102,324. Ordinarily, the liposomes are of the small (from or about 200 to 800 Angstroms) unilamellar type in which the lipid content is greater than about 30 mol percent cholesterol, the selected proportion being adjusted for the most efficacious therapy.

For parenteral administration, in one embodiment cyclic G-2-AllylP is formulated generally by mixing each at the desired degree of purity, in a unit dosage injectable form (solution, suspension, or emulsion), with a pharmaceutically, or parenterally, acceptable carrier, i.e., one that is non-toxic to recipients at the dosages and concentrations employed and is compatible with other ingredients of the formulation.

Generally, the formulations are prepared by contacting cyclic G-2-AllylP uniformly and intimately with liquid carriers or finely divided solid carriers or both. Then, if necessary, the product is shaped into the desired formulation. Preferably the carrier is a parenteral carrier, more preferably a solution that is isotonic with the blood of the recipient. Examples of such carrier vehicles include water, saline, Ringier's solution, a buffered solution, and dextrose solution. Non-aqueous vehicles such as fixed oils and ethyl oleate are also useful herein.

A carrier suitably contains minor amounts of additives such as substances that enhance isotonicity and chemical stability. Such materials are non-toxic to recipients at the dosages and concentrations employed, and include buffers such as phosphate, citrate, succinate, acetic acid, and other organic acids or their salts; antioxidants such as ascorbic acid; low molecular weight (less than about ten residues) polypeptides, e.g., polyarginine or tripeptides; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; glycine; amino acids such as glutamic acid, aspartic acid, histidine, or arginine; monosaccharides, disaccharides, and other carbohydrates including cellulose or its derivatives, glucose, mannose, trehalose, or dextrans; chelating agents such as EDTA; sugar alcohols such as mannitol or sorbitol; counter-ions such as sodium; non-ionic surfactants such as polysorbates, poloxamers, or polyethylene glycol (PEG); and/or neutral salts, e.g., NaCl, KCl, MgCl₂, CaCl₂, etc.

Cyclic G-2-AllylP is typically formulated in such vehicles at a pH of from or about 4.5 to 8. It will be understood that use of certain of the foregoing excipients, carriers, or stabilizers will result in the formation of salts of the compound. The final preparation may be a stable liquid or lyophilized solid.

Formulations of cyclic G-2-AllylP in pharmaceutical compositions can also include adjuvants. Typical adjuvants which may be incorporated into tablets, capsules, and the like are a

binder such as acacia, corn starch, or gelatin; an excipient such as microcrystalline cellulose; a disintegrating agent like corn starch or alginic acid; a lubricant such as magnesium stearate; a sweetening agent such as sucrose or lactose; a flavouring agent such as peppermint, wintergreen, or cherry. When dosage forms are tablets, cyclic G-2-AllylP compositions can include binders and optionally, a smooth coating. When the dosage form is a capsule, in addition to the above materials, it may also contain a liquid carrier such as a fatty oil. Other materials of various types may be used as coatings or as modifiers of the physical form of the dosage unit. A syrup or elixir may contain the active compound, a sweetener such as sucrose, preservatives like propyl paraben, a colouring agent, and a flavouring agent such as cherry. Sterile compositions for injection can be formulated according to conventional pharmaceutical practice. For example, dissolution or suspension of the active compound in a vehicle such as water or naturally occurring vegetable oil like sesame, peanut, or cottonseed oil or a synthetic fatty vehicle like ethyl oleate or the like may be desired. buffers, preservatives, antioxidants, and the like can be incorporated according to accepted pharmaceutical practice.

For injection, intraventricular administration and other invasive routes of administration, cyclic G-2-AllylP must be sterile. Sterility may be accomplished by any method known in the art, for example filtration through sterile filtration membranes (e.g., 0.2 micron membranes). Therapeutic compositions generally are placed into a container having a sterile access port, for example, an intravenous solution bag or vial having a stopper able to be pierced by a hypodermic injection needle.

A pharmaceutical formulation containing cyclic G-2-AllylP ordinarily will be stored in unit or multi-dose containers, for example, in sealed ampoules or vials, as an aqueous solution or as a lyophilized formulation for reconstitution. As an example of a lyophilized formulation, 10 mL vials are filled with 5 mL of sterile-filtered 1% (w/v) aqueous solution of compound, and the resulting mixture is lyophilized. The infusion solution is prepared by reconstituting the lyophilized compound using bacteriostatic Water-for-Injection. It can be readily appreciated that other dosage forms and types of preparations can be used, and all are considered to be part of this invention.

Preparation of the Compounds

Starting materials and reagents used in preparing cyclic G-2-AllylP are either available from commercial suppliers such as Aldrich Chemical Company (Milwaukee, Wis.), Bachem (Torrance, Calif.), Sigma (St. Louis, Mo.), or are prepared by methods well known to the person of ordinary skill in the art following procedures described in such references as Fieser and Fieser's *Reagents for Organic Synthesis*, vols 1-17, John Wiley and Sons, New York, N.Y., 1991; Rodd's *Chemistry of Carbon Compounds*, vols. 1-5 and supplements, Elsevier Science Publishers, 1989; *Organic Reactions*, vols. 1-40, John Wiley and Sons, New York, N.Y., 1991; March J; *Advanced Organic Chemistry*, 4th ed. John Wiley and Sons, New York, N.Y., 1992; and Larock: *Comprehensive Organic Transformations*, VCH Publishers, 1989. In most instances, amino acids and their esters or amides, and protected amino acids, are widely commercially available; and the preparation of modified amino acids and their amides or esters are extensively described in the chemical and biochemical literature and thus well-known to persons of ordinary skill in the art.

Starting materials, intermediates, and final products this invention may be isolated and purified using conventional techniques, including filtration, distillation, crystallization,

chromatography, and the like. They may be characterized using conventional methods, including physical constants and spectral data.

Cyclic G-2-AllylP is a cyclic dipeptide (bicyclic 2,5-diketopiperazine). In general, cyclic G-2-AllylP may be prepared by methods such as are already well-known to persons of ordinary skill in the art of peptide and modified peptide synthesis, following the reaction schemes set forth in the Figures following this specification, or by following other methods well-known to those of ordinary skill in the art of the synthesis of peptides and analogues. See for example, Bodanzsky: *Principles of Peptide Synthesis*, Berlin, New York: Springer-Verlag 1993. Synthesis of the diketopiperazine compounds of this invention may be by solution-phase synthesis as discussed in the Examples or via the solid-phase synthesis method exemplified by Merrifield et al. 1963 *J. Amer. Chem. Soc.*: 85, 2149-2156. Solid phase synthesis may be performed using commercial peptide synthesizers, such as the Applied Biosystems Model 430A, using the protocols established for the instrument.

Specific examples of diketopiperazine synthesis can be found in the Examples following and in, for example, Fischer, 2003, *J. Peptide Science*: 9: 9-35 and references therein. A person of ordinary skill in the art will have no difficulty, taking account of that skill and the knowledge available, and of this disclosure, in developing one or more suitable synthetic methods for compounds of this invention.

The choice of appropriate protecting groups for the method chosen (solid-phase or solution-phase), and of appropriate substrates if solid-phase synthesis is used, will be within the skill of a person of ordinary skill in the art. Appropriate protecting groups for peptide synthesis include t-butyloxycarbonyl (Boc), fluorenylmethyloxycarbonyl (Fmoc), Benzyl (Bzl), t-amylloxycarbonyl (Aoc), tosyl (Tos), benzyloxycarbonyl (Z or Cbz), o-bromo-benzyloxycarbonyl (BrZ) and the like. Additional protecting groups are identified in Goodman M. (ed.), "Synthesis of Peptides and Peptidomimetics" in *Methods of organic chemistry* (Houben-Weyl) (Workbench Edition, E22a,b,c,d,e; 2004; Georg Thieme Verlag, Stuttgart, New York).

The choice of coupling agent for the method chosen will also be within the skill of a person of ordinary skill in the art. Suitable coupling agents include DCC(N,N'-Dicyclohexylcarbodiimide), Bop (Benzotriazole-1-yl-oxy-tris(dimethylamino)-phosphonium hexafluorophosphate), PyBop (Benzotriazole-1-yl-oxytripyrrolidinophosphonium hexafluorophosphate), BopCl (bis(2-oxo-3-oxazolidinyl)phosphinic chloride), 2-Chloro-1,3-dimethylimidazolidinium hexafluorophosphate (CIP) and the like. Other compounds may be used in the synthesis e.g. to prevent racemisation, such as HOBt (N-Hydroxybenzotriazole) and HOAt (1-Hydroxy-7-azabenzotriazole).

All patent and literature references cited throughout the specification are expressly incorporated by reference in their entirety as if each had been separately so incorporated.

EXAMPLES

The present invention is further illustrated by the following examples. These examples are offered by way of illustration only and are not intended to limit the scope of the invention. General Methods

Flash chromatography was performed using Scharlau 60 (40-60 µm mesh) silica gel. Analytical thin layer chromatography was carried out on 0.20 mm pre-coated silica gel plates (ALUGRAM® SIL G/UV₂₅₄) and compounds visualized

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using UV fluorescence, or heating of plates dipped in potassium permanganate in alkaline solution.

Melting points in degrees Celsius ($^{\circ}\text{C}.$) were determined on an Electrothermal[®] melting point apparatus and are uncorrected.

Optical rotations were measured at $20^{\circ}\text{C}.$ on a Perkin Elmer 341 polarimeter using 10 cm path length cells and are given in units of $10^{-1}\text{ deg cm}^2\text{ g}^{-1}$. Samples were prepared in the solvent indicated at the concentration specified (measured in $\text{g}/100\text{ cm}^3$). IR spectra were recorded on a Perkin Elmer Spectrum One FT-IR spectrometer. The samples were prepared as thin films on sodium chloride discs or as solids in potassium bromide discs. A broad signal indicated by br. The frequencies (ν) as absorption maxima are given in wavenumbers (cm^{-1}).

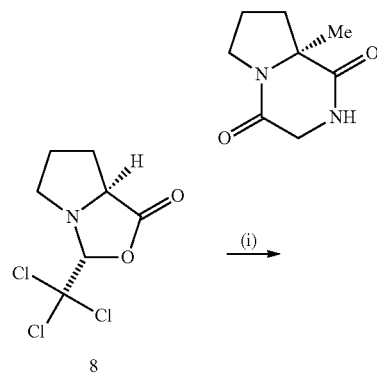
NMR spectra were recorded on a Bruker AVANCE DRX400 (^1H , 400 MHz; ^{13}C , 100 MHz) or a Bruker AVANCE 300 (^1H , 300 MHz; ^{13}C , 75 MHz) spectrometer at ambient temperatures. For ^1H NMR data chemical shifts are described in parts per million downfield from SiMe_4 and are reported consecutively as position (δ_H), relative integral, multiplicity (s=singlet, d=doublet, t=triplet, dd=doublet of doublets, m=multiplet, br=broad), coupling constant (J/Hz) and assignment. For ^{13}C NMR data, chemical shifts are described in parts per million relative to CDCl_3 and are reported consecutively as position (δ_C), degree of hybridization as determined by DEPT experiments, and assignment. ^1H NMR spectra were referenced internally using SiMe_4 (δ 0.00) or CDCl_3 (δ 7.26). ^{13}C NMR spectra were referenced internally using CDCl_3 (δ 77.0). When two sets of peaks arise in the NMR spectra due to different conformations around the glycine-proline amide bond, the chemical shift for the minor cis conformer is marked with an asterisk (*).

Accurate mass measurements were recorded on a VG-70SE mass spectrometer. Hexane and dichloromethane were distilled prior to use. Methanol was dried using magnesium turnings and iodine, and distilled under nitrogen. Triethylamine was dried over calcium hydride and distilled under nitrogen.

Example 1

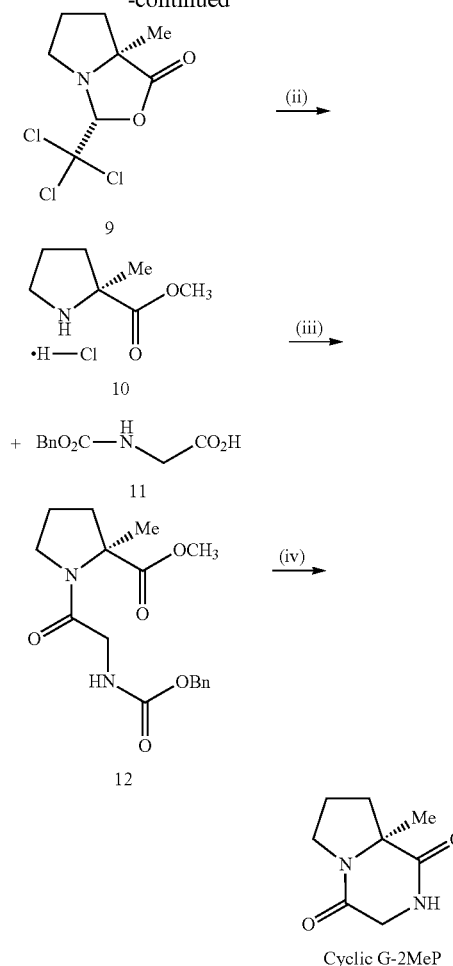
Synthesis of (8aS)-Methyl-hexahydropyrrolo[1,2-a]pyrazine-1,4-dione (Cyclic G-2MeP)

Scheme 1: Reagents, conditions and yields:



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-continued



(i) LDA, THF, $-78^{\circ}\text{C}.$, iodomethane, $-78 \rightarrow -50^{\circ}\text{C}.$, 2 h (63%); (ii) SOCl_2 , CH_3OH , reflux, N_2 , 2.5 h (98%); (iii) Et_3N , BocCl , CH_2Cl_2 , RT, N_2 , 20.5 h (78%); (iv) 10% Pd/C , CH_3OH , RT, 15 h (98%).

(2R,5S)-4-Methyl-2-trichloromethyl-1-aza-3-oxabicyclo[3.3.0]octan-4-one 9

$n\text{-BuLi}$ (1.31 M, 4.68 cm^3 , 6.14 mmol) was added dropwise to a stirred solution of diisopropylamine (0.86 cm^3 , 6.14 mmol) in dry tetrahydrofuran (10 cm^3) at $-78^{\circ}\text{C}.$ under an atmosphere of nitrogen. The solution was stirred for 5 min, warmed to $0^{\circ}\text{C}.$ and stirred for 15 min. The solution was then added dropwise to a solution of oxazolidinone 8 (1.00 g, 4.09 mmol) in dry tetrahydrofuran (20 cm^3) at $-78^{\circ}\text{C}.$ over 20 min (turned to a dark brown colour), stirred for a further 30 min then iodomethane (0.76 cm^3 , 12.3 mmol) was added dropwise over 5 min. The solution was warmed to $-50^{\circ}\text{C}.$ over 2 h. Water (15 cm^3) was added and the solution warmed to room temperature and extracted with chloroform ($3 \times 40\text{ cm}^3$). The combined organic extracts were dried (MgSO_4), filtered and evaporated to dryness in vacuo to give a dark brown semi-solid. Purification of the residue by flash column chromatography (15% ethyl acetate-hexane) afforded oxazolidinone 9 (0.67 g, 63%) as a pale yellow solid: mp $55\text{--}57^{\circ}\text{C}.$ (lit., $57\text{--}60^{\circ}\text{C}.$); δ_H (300 MHz, CDCl_3) 1.53 (3H, s, CH_3), 1.72–2.02 (3H, m, $\text{Pro}\beta\text{-H}$ and $\text{Pro}\gamma\text{-H}_2$), 2.18–2.26 (1H, m, $\text{Pro}\beta\text{-H}$), 3.15–3.22 (1H, m, $\text{Pro}\delta\text{-H}$), 3.35–3.44 (1H, m, $\text{Pro}\delta\text{-H}$) and 4.99 (1H, s, NCH).

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Methyl L-2-methylproline hydrochloride 10

a) Using Acetyl Chloride

Oxazolidinone 9 (0.60 g, 2.33 mmol) was dissolved in dry methanol (15 cm³) under an atmosphere of nitrogen and acetyl chloride (0.33 cm³, 4.66 mmol) was added dropwise to the ice-cooled solution. The solution was heated under reflux for 4.5 h, then the solvent removed under reduced pressure to give a brown oil which was purified by flash column chromatography (10% CH₃OH—CH₂Cl₂) affording the hydrochloride 10 (0.2 g, 48%) as a flaky white solid: mp 107–109° C. (lit., 106–108° C.); δ_H (300 MHz, CDCl₃) 1.81 (3H, s, CH₃), 1.93–2.14 (3H, m, Pro β -H_AH_B and Pro γ -H₂), 2.33–2.39 (1H, m, Pro β -H_AH_B), 3.52–3.56 (2H, m, Pro δ -H₂) and 3.82 (3H, s, CO₂CH₃).

b) Using Thionyl Chloride

An ice-cooled solution of oxazolidinone 9 (53 mg, 0.21 mmol) in dry methanol (1 cm³) was treated dropwise with thionyl chloride (0.045 cm³, 0.62 mmol). The solution was heated under reflux for 2.5 h, cooled and the solvent removed under reduced pressure to yield a brown oil. The oil was dissolved in toluene (5 cm³), concentrated to dryness to remove residual thionyl chloride and methanol then purified by flash column chromatography (10% CH₃OH—CH₂Cl₂) to afford the hydrochloride 10 (16 mg, 43%) as a flaky white solid. The ¹H NMR assignments were in agreement with those reported above.

Methyl-N-benzoyloxycarbonyl-glycyl-L-2-methylproline 12

Dry triethylamine (0.27 cm³, 1.96 mmol) was added dropwise to a solution of hydrochloride 10 (0.11 g, 0.61 mmol) and N-benzoyloxycarbonyl-glycine 11 (98.5%) (0.17 g, 0.79 mmol) in dry dichloromethane (35 cm³) under an atmosphere of nitrogen at room temperature, and the reaction mixture stirred for 10 min. Bis(2-oxo-3-oxazolidinyl)phosphinic chloride (BoPCI, 97%) (0.196 g, 0.77 mmol) was added and the resultant colourless solution was stirred for 20.5 h. The solution was washed successively with 10% aqueous hydrochloric acid (30 cm³) and saturated aqueous sodium hydrogen carbonate (30 cm³), dried (MgSO₄), filtered and evaporated to dryness in vacuo. Purification of the resultant residue by flash column chromatography (50–80% ethyl acetate–hexane; gradient elution) yielded dipeptide 12 (0.18 g, 92%) as a colourless oil. Amide 12 was shown to exist as a 98:2 trans:cis mixture of conformers by ¹³C NMR analysis (the ratio was estimated from the relative intensities of the resonances at δ 20.8 and 23.5 assigned to the Pro γ -C atoms of the minor and major conformers, respectively): [α]_D –33.0 (c 1.0 in MeOH); ν_{max} (film)/cm^{–1} 3406, 2952, 1732, 1651, 1521, 1434, 1373, 1329, 1310, 1284, 1257, 1220, 1195, 1172, 1135, 1107, 1082, 1052, 1029, 986, 965, 907, 876, 829, 775, 738 and 699; δ_H (300 MHz, CDCl₃) 1.49 (3H, s, CH₃), 1.77–2.11 (4H, m, Pro β -H₂ and Pro γ -H₂), 3.43–3.48 (2H, m, Pro δ -H₂), 3.61 (3H, s, OCH₃), 3.85–3.89 (2H, m, Gly α -H₂), 5.04 (2H, s, PhCH₂), 5.76 (1H, br s, N—H) and 7.21–7.28 (5H, s, ArH); δ_C (75 MHz, CDCl₃) 13.8* (CH₃, Pro α -CH₃), 21.1 (CH₃, Pro α -CH₃), 20.8* (CH₂, Pro γ -C), 23.5 (CH₂, Pro γ -C), 38.0 (CH₂, Pro β -C), 40.8* (CH₂, Pro β -C), 43.3 (CH₂, Gly α -C), 45.5* (CH₂, Gly α -C), 46.6 (CH₂, Pro δ -C), 48.7* (CH₂, Pro δ -C), 51.9* (CH₃, OCH₃), 52.1 (CH₃, OCH₃), 60.0* (quat., Pro α -C), 66.0 (quat., Pro α -C), 66.3 (CH₂, PhCH₂), 68.6* (CH₂, PhCH₂), 127.5 (CH, Ph), 127.6 (CH, Ph), 127.9* (CH, Ph), 128.1 (CH, Ph), 128.3* (CH, Ph), 136.2 (quat., Ph), 155.9 (quat., NCO₂), 166.0 (quat., Gly-CON), 169.4* (quat., Gly-

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CON) and 173.6 (quat., CO₂CH₃); m/z (EI+) 334.1535 (M⁺. C₁₇H₂₂N₂O₅ requires 334.1529).

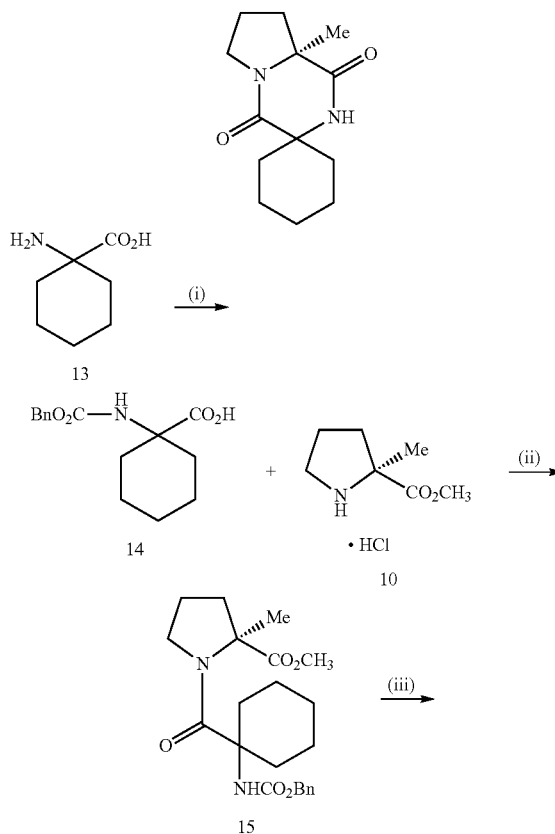
(8aS)-Methyl-hexahydropyrrolo[1,2-a]pyrazine-1,4-dione (Cyclic G-2-MeP)

To a solution of dipeptide 12 (0.167 g, 0.51 mmol) in methanol (8.0 cm³) was added 10% Pd on activated charcoal (8.1 mg, 0.076 mmol) and the vessel flushed with hydrogen gas. The resulting suspension was stirred vigorously under an atmosphere of hydrogen for 15 h. The mixture was then filtered through a Celite pad then a short plug of silica gel with methanol, and the solvent removed under reduced pressure to produce cyclic G-2MeP (83 mg, 98%) as a yellow solid: mp 133–135° C.; [α]_D –128.1 (c 0.52 in MeOH); δ_H (300 MHz, CDCl₃) 1.36 (3H, s, CH₃), 1.87–2.01 (3H, m, Pro β -H_AH_B and Pro γ -H₂), 2.07–2.21 (1H, m, Pro β -H_AH_B), 3.45–3.64 (2H, m, Pro δ -H₂), 3.82 (1H, dd, J 17.1 and 4.1, CH₄H_BNH), 3.99 (1H, d, J 17.1, CH₄H_BNH) and 7.66 (1H, br s, N—H); δ_C (75 MHz, CDCl₃) 20.2 (CH₂, Pro γ -C), 23.2 (CH₃, Pro α -CH₃), 35.0 (CH₂, Pro β -C), 44.7 (CH₂, Pro δ -C), 45.9 (CH₂, CH₂NH), 63.8 (quat., Pro α -C), 163.3 (quat., NCO) and 173.3 (quat., CONH); m/z (EI+) 168.08986 (M⁺. C₈H₁₂N₂O₂ requires 168.08988).

Example 2

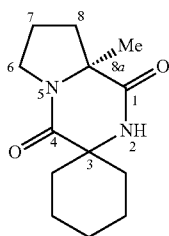
Synthesis of (8aS)-Methyl-spiro[cyclohexane-1,3-(4H)-tetrahydropyrrolo[1,2-a]pyrazine]-1,4(2H)-dione (Cyclic cyclohexyl-G-2-MeP)

Scheme 2: Reagents, conditions and yields:



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-continued



Cyclic cyclohexyl-G-2-MeP

(i) BnO_2CCl , Na_2CO_3 , H_2O -dioxane (3:1), 19 h, 96%; (ii) Et_3N , HOAt , CIP , 1,2-dichloroethane, reflux, N_2 , 19 h (23%); (iii) 10% Pd/C , CH_3OH , RT, 17 h (65%).

N-Benzylloxycarbonyl-1-aminocyclohexane-1-carboxylic acid (14)

To a suspension of 1-aminocyclohexanecarboxylic acid 13 (0.72 g, 5.02 mmol) and sodium carbonate (1.6 g, 15.1 mmol) were dissolved in water-dioxane (21 cm^3 , 3:1) was added benzyl chloroformate (0.79 cm^3 , 5.52 mmol) was added dropwise and the solution was stirred at room temperature for 19.5 h. The aqueous layer was washed with diethyl ether (60 cm^3), acidified with 2 M HCl and extracted with ethyl acetate (2 \times 60 cm^3). The organic layers were combined, dried (MgSO_4), filtered and evaporated under reduced pressure to produce a colourless oil, which solidified on standing to crude carbamate 14 (1.23 g, 88%) as a white solid: mp 152-154° C. (lit., 148-150° C.); δ_{H} (400 MHz, CDCl_3) 1.27-1.56 (3H, m, 3 \times cyclohexyl-H), 1.59-1.73 (3H, m, 3 \times cyclohexyl-H), 1.85-1.91 (2H, m, 2 \times cyclopentyl-H), 2.05-2.09 (2H, m, 2 \times cyclopentyl-H), 5.02 (1H, br s, N—H), 5.12 (2H, s, OCH_2Ph) and 7.27-7.36 (5H, s, Ph); δ_{C} (100 MHz, CDCl_3) 21.1 (CH_2 , 2 \times cyclohexyl-C), 25.1 (CH_2 , 2 \times cyclohexyl-C), 32.3 (CH_2 , cyclohexyl-C), 59.0 (quat., 1-C), 67.1 (CH_2 , OCH_2Ph), 128.1 (CH, Ph), 128.2 (CH, Ph), 128.5 (CH, Ph), 136.1 (quat., Ph), 155.7 (quat., NCO_2) and 178.7 (quat., CO_2H).

Methyl-N-benzylloxycarbonyl-cyclohexyl-glycyl-L-2-methylproline (15)

Dry triethylamine (0.21 cm^3 , 1.5 mmol) was added dropwise to a solution of hydrochloride 10 (84.0 mg, 0.47 mmol), carboxylic acid 14 (0.17 g, 0.61 mmol) and 1-hydroxy-7-azabenzotriazole (16 mg, 0.12 mmol) in dry 1,2-dichloroethane (26 cm^3) under an atmosphere of nitrogen at room temperature, and the reaction mixture stirred for 10 min. 2-Chloro-1,3-dimethylimidazolidinium hexafluorophosphate (0.13 g, 0.47 mmol) was added and the resultant solution heated under reflux for 21 h, then washed successively with 10% aqueous hydrochloric acid (30 cm^3) and saturated aqueous sodium hydrogen carbonate (30 cm^3), dried (MgSO_4), filtered and evaporated to dryness in vacuo. Purification of the resultant residue by flash column chromatography (40-50% ethyl acetate-hexane; gradient elution) yielded amide 15 (16 mg, 9%) as a white solid. Amide 15 was shown to exist as a 11:1 trans:cis mixture of conformers by ^{13}C NMR analysis (the ratio was estimated from the relative intensities of the resonances at δ 41.3 and 48.2 assigned to the Pro δ -C atoms of the minor and major conformers, respectively): mp 219-222° C.; $[\alpha]_{\text{D}} -44.9$ (c 1.31 in CH_2Cl_2); ν_{max} (film)/ cm^{-1} 3239, 2927, 1736, 1707, 1617, 1530, 1450, 1403, 1371, 1281, 1241, 1208, 1194, 1165, 1150, 1132, 1089, 1071, 1028, 984, 912, 796, 749, 739 and 699; δ_{H} (400 MHz, CDCl_3) 1.24-2.10 (17H, m, Pro α - CH_3 , Pro β - H_2 , Pro γ - H_2 and 5 \times

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cyclohexyl- H_2), 3.25-3.48 (1H, br m, Pro δ - H_AH_B), 3.61-3.87 (4H, br m, OCH_3 and Pro δ - H_AH_B), 4.92-5.19 (3H, m, N—H and OCH_2Ph) and 7.35-7.37 (5H, s, Ph); δ_{C} (100 MHz, CDCl_3) 21.26 (CH_2 , cyclohexyl-C), 21.33 (CH_2 , cyclohexyl-C), 21.7 (CH_3 , Pro α - CH_3), 24.8 (CH_2 , cyclohexyl-C), 25.0 (CH_2 , Pro γ -C), 29.4* (CH_2 , cyclohexyl-C), 29.7* (CH_2 , cyclohexyl-C), 31.1 (CH_2 , cyclohexyl-C), 31.6 (CH_2 , cyclohexyl-C), 31.9* (CH_2 , cyclohexyl-C), 32.2* (CH_2 , cyclohexyl-C), 32.8* (CH_2 , cyclohexyl-C), 37.3 (CH_2 , Pro β -C), 41.4* (CH_2 , Pro δ -C), 48.2 (CH_2 , Pro δ -C), 52.1 (CH_3 , OCH_3), 59.1 (quat., Gly α -C), 66.7 (CH_2 , OCH_2Ph), 67.3* (CH_2 , OCH_2Ph), 67.4 (quat., Pro α -C), 128.0* (CH, Ph), 128.1* (CH, Ph), 128.3 (CH, Ph), 128.5 (CH, Ph), 128.7 (CH, Ph), 136.6 (quat., Ph), 153.7 (quat., NCO_2), 171.0 (quat., Gly-CO) and 174.8 (quat., CO_2CH_3); m/z (EI+) 402.2151 (M^+ . $\text{C}_{22}\text{H}_{30}\text{N}_2\text{O}_5$ requires 402.2155).

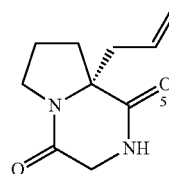
(8aS)-Methyl-spiro[cyclohexane-1,3(4H)-tetrahydropyrrolo[1,2-c]pyrazine]-1,4(2H)-dione (Cyclic cyclohexyl-G-2-MeP)

To a solution of amide 15 (40 mg, 0.01 mmol) in methanol (3.3 cm^3) was added 10% Pd on activated charcoal (1.6 mg, 0.015 mmol) and the vessel flushed with hydrogen gas. The resulting suspension was stirred vigorously under an atmosphere of hydrogen for 61.5 h, then filtered through a CeliteTM pad with methanol (15 cm^3). The filtrate was concentrated to dryness under reduced pressure to produce a yellow semi-solid which was purified by reverse-phase C18 flash column chromatography (0-10% $\text{CH}_3\text{CN}/\text{H}_2\text{O}$; gradient elution) to produce cyclic cyclohexyl-G-2-MeP (19 mg, 81%) as a white solid: mp 174-177° C.; $[\alpha]_{\text{D}} -63.8$ (c 1.13 in CH_2Cl_2); ν_{max} (film)/ cm^{-1} 3215, 2925, 2854, 1667, 1646, 1463, 1427, 1276, 1232, 1171, 1085, 1014, 900, 868, 818, 783, 726 and 715; δ_{H} (400 MHz, CDCl_3) 1.31-1.89 (12H, m, 9 \times cyclohexyl-H and 8a- CH_3), 1.94-2.15 (4H, m, 7- H_2 and 8- H_2), 2.26 (1H, td, J 13.7 and 4.5, 1 \times cyclohexyl-H), 3.44-3.51 (1H, m, 3.79-3.86 (1H, m, 6- H_AH_B) and 6.40 (1H, br s, N—H); δ_{C} (100 MHz, CDCl_3) 19.5 (CH_2 , 7-C), 20.6 (CH_2 , cyclohexyl-C), 20.8 (CH_2 , cyclohexyl-C), 24.5 (CH_2 , cyclohexyl-C), 25.0 (CH_3 , 8a- CH_3), 33.7 (CH_2 , cyclohexyl-C), 36.3 (CH_2 , 8-C), 36.5 (CH_2 , cyclohexyl-C), 44.7 (CH_2 , 6-C), 59.5 (quat., 8a-C), 64.0 (quat., 3-C), 168.1 (quat., 4-C) and 171.6 (quat., 1-C); m/z (EI+) 236.15246 (M^+ . $\text{C}_{13}\text{H}_{20}\text{N}_2\text{O}_2$ requires 236.15248).

Example 3

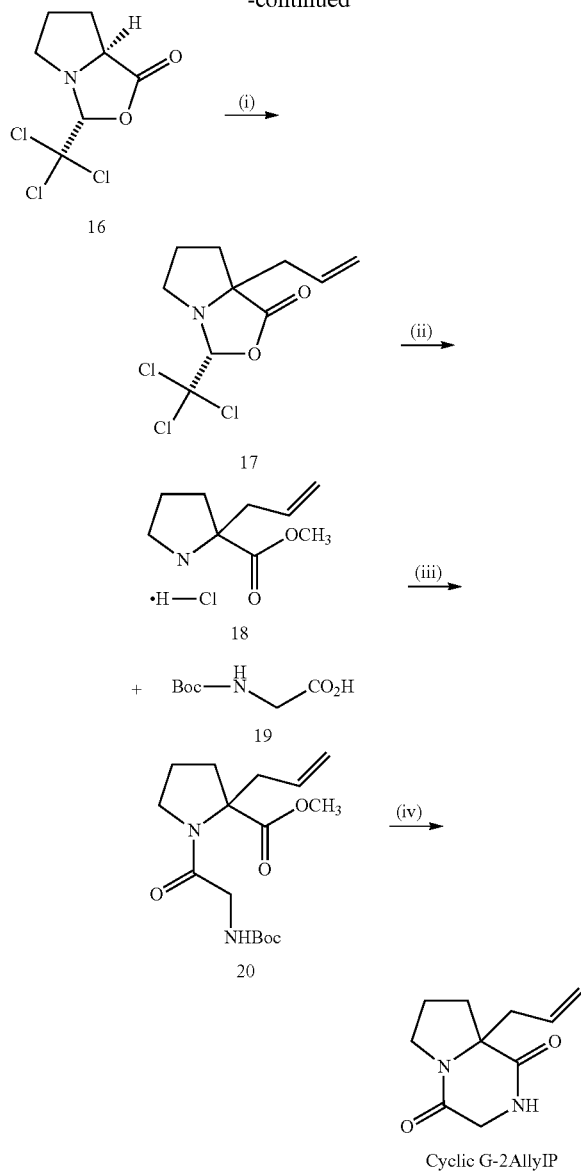
Synthesis of (8aS)-Allyl-hexahydropyrrolo[1,2-a]pyrazine-1,4-dione (Cyclic G-2-AllylP)

Scheme 3: Reagents, conditions and yields:



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-continued



(i) LDA, THF, -78°C ., allyl bromide, $-78 \rightarrow -30^{\circ}\text{C}$., N_2 , 4 h (60%); (ii) acetyl chloride, CH_3OH , reflux, N_2 , 24 h (63%); (iii) Et_3N , BoPCl , CH_2Cl_2 , RT, N_2 , 19.5 h (45%); (iv) TFA, CH_2Cl_2 , 1 h, then Et_3N , CH_2Cl_2 , 23 h (37%).

(2R,5S)-4-Allyl-2-trichloromethyl-1-aza-3-oxabicyclo[3.3.0]octan-4-one 17

$n\text{-BuLi}$ (1.31 M, 9.93 cm^3 , 13.0 mmol) was added dropwise to a stirred solution of diisopropylamine (1.82 cm^3 , 13.0 mmol) in dry tetrahydrofuran (20 cm^3) at -78°C . under an atmosphere of nitrogen. The solution was stirred for 5 min, warmed to 0°C ., stirred for 15 min then added dropwise to a solution of pro-oxazolidinone 16 (2.12 g, 8.68 mmol) in dry tetrahydrofuran (40 cm^3) at -78°C . over 20 min and the reaction mixture was stirred for a further 30 min then allyl bromide (2.25 cm^3 , 26.0 mmol) was added dropwise over 5 min. The solution was warmed slowly to -30°C . over 4 h, quenched with H_2O (30 cm^3) and the mixture warmed to room temperature and extracted with chloroform ($3 \times 80\text{ cm}^3$). The combined organic extracts were dried (MgSO_4), filtered and evaporated to dryness in vacuo to produce a dark brown

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semi-solid which was purified by flash column chromatography (10-20% ethyl acetate-hexane; gradient elution) to produce oxazolidinone 17 (1.48 g, 60%) as an orange oil which solidified at 0°C ., for which the nmr data were in agreement with that reported in the literature: δ_{H} (400 MHz, CDCl_3) 1.58-1.92 (2H, m, $\text{Pro}\gamma\text{-H}_2$), 1.96-2.14 (2H, m, $\text{Pro}\beta\text{-H}_2$), 2.50-2.63 (2H, m, $\text{Pro}\delta\text{-H}_2$), 3.12-3.23 (2H, m, $\text{CH}_2\text{CH}=\text{CH}_2$), 4.97 (1H, s, NCH), 5.13-5.18 (2H, m, $\text{CH}=\text{CH}_2$) and 5.82-5.92 (1H, m, $\text{CH}=\text{CH}_2$); δ_{C} (100 MHz, CDCl_3) 25.1 (CH_2 , $\text{Pro}\gamma\text{-C}$), 35.1 (CH_2 , $\text{Pro}\beta\text{-C}$), 41.5 (CH_2 , $\text{Pro}\delta\text{-C}$), 58.3 (CH_2 , $\text{CH}_2\text{CH}=\text{CH}_2$), 71.2 (quat., $\text{Pro}\alpha\text{-C}$), 100.4 (quat., CCl_3), 102.3 (CH, NCH), 119.8 (CH_2 , $\text{CH}_2\text{CH}=\text{CH}_2$), 131.9 (CH, $\text{CH}_2\text{CH}=\text{CH}_2$) and 176.1 (quat., $\text{C}=\text{O}$); m/z (CI^+) 284.0009 [($\text{M}+\text{H}$) $^+$. $\text{C}_{10}\text{H}_{13}^{35}\text{Cl}_3\text{NO}_2$ requires 284.0012], 285.9980 [($\text{M}+\text{H}$) $^+$. $\text{C}_{10}\text{H}_{13}^{35}\text{Cl}_2^{37}\text{ClNO}_2$ requires 285.9982], 287.9951 [($\text{M}+\text{H}$) $^+$. $\text{C}_{10}\text{H}_{13}^{35}\text{Cl}^{37}\text{Cl}_2\text{NO}_2$ requires 287.9953] and 289.9932 [($\text{M}+\text{H}$) $^+$. $\text{C}_{10}\text{H}_{13}^{37}\text{Cl}_3\text{NO}_2$ requires 289.9923].

Methyl L-2-allylproline hydrochloride 18

An ice-cooled solution of oxazolidinone 17 (0.64 g, 2.24 mmol) in dry methanol (15 cm^3) was treated dropwise with a solution of acetyl chloride (0.36 cm^3 , 5.0 mmol) in methanol (5 cm^3). The solution was heated under reflux for 24 h, then cooled and the solvent removed under reduced pressure. The resultant brown oil was dissolved in toluene (40 cm^3) and concentrated to dryness to remove residual thionyl chloride and methanol, then purified by flash column chromatography (5-10% $\text{CH}_3\text{OH}-\text{CH}_2\text{Cl}_2$; gradient elution) to afford hydrochloride 18 (0.29 g, 63%) as a green solid for which the NMR data were in agreement with that reported in the literature: δ_{H} (300 MHz, CDCl_3) 1.72-2.25 (3H, m, $\text{Pro}\beta\text{-H}_A\text{H}_B$ and $\text{Pro}\gamma\text{-H}_2$), 2.32-2.52 (1H, m, $\text{Pro}\beta\text{-H}_A\text{H}_B$), 2.72-3.10 (2H, m, $\text{Pro}\delta\text{-H}_2$), 3.31-3.78 (2H, m, $\text{CH}_2\text{CH}=\text{CH}_2$), 3.84 (3-1, s, CO_2CH_3), 5.20-5.33 (2H, m, $\text{CH}=\text{CH}_2$), 5.75-5.98 (1H, m, $\text{CH}=\text{CH}_2$) and 8.06 (1H, br s, N—H); m/z (CI^+) 170.1183 [($\text{M}+\text{H}$) $^+$. $\text{C}_9\text{H}_{16}\text{NO}_2$ requires 170.1181].

Methyl-N-tert-butyloxycarbonyl-glycyl-L-2-allylproline 20

Dry triethylamine (0.28 cm^3 , 2.02 mmol) was added dropwise to a solution of hydrochloride 18 (0.13 g, 0.63 mmol) and N-tert-butyloxycarbonyl-glycine 19 (0.14 g, 0.82 mmol) in dry dichloromethane (35 cm^3) under an atmosphere of nitrogen at room temperature, and the reaction mixture was stirred for 10 min. Bis(2-oxo-3-oxazolidinyl)phosphinic chloride (BoPCl , 97%) (0.20 g , 0.80 mmol) was added and the solution stirred for 19.5 h, then washed successively with 10% aqueous hydrochloric acid (35 cm^3) and saturated aqueous sodium hydrogen carbonate (35 cm^3), dried (MgSO_4), filtered and evaporated to dryness in vacuo. Purification of the resultant residue by flash column chromatography (40% ethyl acetate-hexane) yielded dipeptide 20 (0.09 g, 45%) as a light yellow oil: $[\alpha]_{\text{D}}^{25} +33.8$ (c 0.83 in CH_2Cl_2); ν_{max} (film)/ cm^{-1} 3419, 3075, 2977, 2930, 2874, 1739, 1715, 1656, 1499, 1434, 1392, 1366, 1332, 1268, 1248, 1212, 1168, 1122, 1051, 1026, 1003, 943, 919, 867, 830, 779, 739, 699 and 679; δ_{H} (300 MHz, CDCl_3) 1.42 [9H, s, $\text{C}(\text{CH}_3)_3$], 1.93-2.08 (4H, m, $\text{Pro}\beta\text{-H}_2$ and $\text{Pro}\gamma\text{-H}_2$), 2.59-2.67 (1H, m, $\text{CH}_A\text{H}_B\text{CH}=\text{CH}_2$), 3.09-3.16 (1H, m, $\text{CH}_A\text{H}_B\text{CH}=\text{CH}_2$), 3.35-3.44 (1H, m, $\text{Pro}\delta\text{-H}_A\text{H}_B$), 3.56-3.62 (1H, m, $\text{Pro}\delta\text{-H}_A\text{H}_B$), 3.70 (3H, s, OCH_3), 3.89 (2H, d, J 4.2, $\text{Gly}\alpha\text{-H}_2$), 5.06-5.11 (2H, m, $\text{CH}=\text{CH}_2$), 5.42 (1H, br s, Gly-NH) and 5.58-5.72 (1H, m, $\text{CH}=\text{CH}_2$); δ_{C} (75 MHz, CDCl_3) 23.7 (CH_2 , $\text{Pro}\gamma\text{-C}$), 28.3 [CH_3 , $\text{C}(\text{CH}_3)_3$], 35.0 (CH_2 , $\text{Pro}\beta\text{-C}$), 37.6 (CH_2 , $\text{CH}_2\text{CH}=\text{CH}_2$),

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43.3 (CH₂, Glyα-C), 47.5 (CH₂, Proδ-C), 52.5 (CH₃, OCH₃), 68.8 (quat., Proα-C), 79.5 [quat., C(CH₃)₃], 119.4 (CH₂, CH=CH₂), 132.9 (CH, CH=CH₂), 155.7 (quat., NCO₂), 166.9 (quat., Gly-CON) and 173.8 (quat., CO₂CH₃); m/z (EI+) 326.1845 (M⁺. C₁₆H₂₆N₂O₅ requires 326.1842).

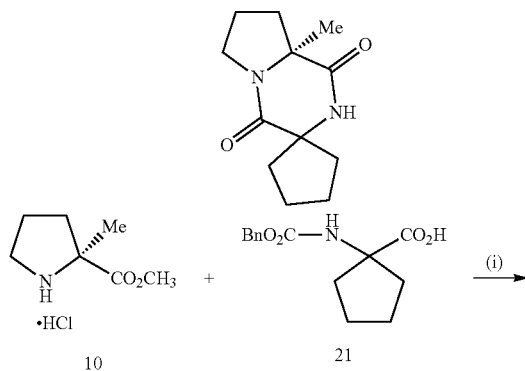
(8aS)-Allyl-hexahydropyrrolo[7,2-a]pyrazine-1,4-dione (Cyclic G-2AllylP)

To a solution of dipeptide 20 (0.09 g, 0.28 mmol) in dichloromethane (9 cm³) at room temperature was added trifluoroacetic acid (1 cm³, 0.013 mmol) dropwise and the reaction mixture was stirred for 1 h under an atmosphere of nitrogen. The solution was evaporated under reduced pressure to give a colorless oil which was dissolved in dichloromethane (10 cm³), dry triethylamine (0.096 cm³, 0.69 mmol) was added and the reaction mixture stirred for 4.5 h, after which further triethylamine (0.096 cm³, 0.69 mmol) was added. The reaction mixture was stirred overnight, concentrated to dryness to give a green oil which was purified by flash column chromatography (10% CH₃OH—CH₂Cl₂) to produce cyclic G-2AllylP (20 mg, 37%) as an off-white solid: mp 106-109° C.; [α]_D -1017 (c 0.95 in CH₂Cl₂); ν_{max} (CH₂Cl₂)/cm⁻¹ 3456, 3226, 2920, 1666, 1454, 1325, 1306, 1299, 1210, 1133, 1109, 1028, 1010, 949, 928, 882, 793, 761 and 733; δ_H (400 MHz, CDCl₃) 1.92-2.01 (2H, m, Proγ-H₂), 1.09-2.16 (2H, m, Proβ-H₂), 2.39-2.56 (2H, m, CH₂CH₂=CH₂), 3.46-3.53 (1H, m, Proδ-H_AH_B), 3.78-3.87 (2H, m, Proδ-H_AH_B and Glyα-H_AH_B), 4.09 (1H, d, J 17.2, Glyα-H_AH_B), 5.16-5.20 (2H, m, CH=CH₂), 5.73-5.84 (1H, m, CH=CH₂) and 7.17 (1H, br s, N—H); δ_C (100 MHz, CDCl₃) 20.1 (CH₂, Proγ-C), 34.1 (CH₂, Proβ-C), 41.7 (CH₂, CH₂CH₂=CH₂), 44.9 (CH₂, Proγ-C), 46.4 (CH₂, Glyα-C), 67.2 (quat., Proα-C), 120.9 (CH₂, CH=CH₂), 131.0 (CH, CH=CH₂), 163.4 (quat., NCO) and 171.7 (quat., CONH); m/z (EI+) 195.1132 (M⁺. C₁₀H₁₅N₂O₂ requires 195.1134).

Example 4

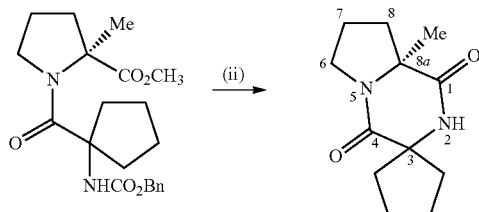
Synthesis of (8aS)-Methyl-spiro[cyclopentane-1,3(4H)-tetrahydropyrrolo[1,2-a]pyrazine]-1,4(2H)-dione (Cyclic Cyclopentyl-G-2-MeP)

Scheme 4: Reagents, conditions and yields: (i) Et₃N, HOAt, CIP, 1,2-dichloroethane, 83° C., N₂, 19 h (23%); (ii) 10% Pd/C, CH₃OH, RT, 17 h (65%).



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-continued



Cyclic cyclopentyl-G-2MeP

N-Benzyloxycarbonyl-L-aminocyclopentane-1-carboxylic acid 21

A solution of benzyl chloroformate (0.290 g, 1.1 mmol) in dioxane (2.5 cm³) was added dropwise to a solution of 1-aminocyclopentanecarboxylic acid (Fluka) (0.2 g, 1.54 mmol) and sodium carbonate (0.490 g, 4.64 mmol) in water (5 cm³) at 0° C. Stirring was continued at room temperature overnight and the reaction mixture washed with ether. The aqueous layer was acidified with 2M hydrochloric acid, extracted with ethyl acetate, dried (Na₂SO₄), filtered and the solvent removed to afford carbamate 21 (0.253 g, 62%) as an oil which solidified on standing. Carbamate 21 was shown to be a 70:30 mixture of conformers by ¹H NMR analysis (the ratio was estimated from the integration of the resonances at δ 5.31 and 7.29-7.40, assigned to the N—H protons of the major and minor conformers, respectively): mp 70-80° C. (lit.¹ 82-86° C., ethyl acetate, petroleum ether); δ_H (400 MHz; CDCl₃; Me₄Si) 1.83 (4H, br s, 2× cyclopentyl-H₂), 2.04 (2H, br s, cyclopentyl-H₂), 2.20-2.40 (2H, m, cyclopentyl-H₂), 5.13 (2H, br s, OCH₂Ph), 5.31 (0.7H, br s, N—H) and 7.29-7.40 (5.3H, m, Ph and N—H*); δ_C (100 MHz; CDCl₃) 24.6 (CH₂, cyclopentyl-C), 37.5 (CH₂, cyclopentyl-C), 66.0 (quat., cyclopentyl-C), 66.8 (CH₂, OCH₂Ph), 128.0 (CH, Ph), 128.1 (CH, Ph), 128.4 (CH, Ph), 136.1 (quat, Ph), 155.8 (quat., NCO₂) and 179.5 (quat., CO₂H).

* denotes resonance assigned to minor conformer.

Methyl N-benzyloxycarbonyl cyclopentyl-glycyl-L-2-methylproline 22

Dry triethylamine (0.19 cm³, 1.4 mmol) was added dropwise to a solution of hydrochloride 10 (78 mg, 0.43 mmol), carboxylic acid 21 (0.15 g, 0.56 mmol) and 1-hydroxy-7-azabenzotriazole (Acros) (15 mg, 0.11 mmol) in dry 1,2-dichloroethane (24 cm³) under an atmosphere of nitrogen at room temperature, and the reaction mixture stirred for 10 min. 2-Chloro-1,3-dimethylimidazolidinium hexafluorophosphate (CIP) (Aldrich) (0.12 g, 0.43 mmol) was added and the resultant solution heated under reflux for 19 h, then washed successively with 10% aqueous hydrochloric acid (30 cm³) and saturated aqueous sodium hydrogen carbonate (30 cm³), dried (MgSO₄), filtered and evaporated to dryness in vacuo. Purification of the resultant residue by flash column chromatography (60% ethyl acetate-hexane) yielded amide 22 (39 mg, 23%) as a white solid. Amide 22 was shown to exist as a 3:1 trans:cis mixture of carbamate conformers by ¹³C NMR analysis (the ratio was estimated from the relative intensities of the resonances at δ 154.1 and 155.7 assigned to the carbamate carbonyl-C atoms of the major and minor conformers, respectively): mp 200-203° C.; [α]_D -54.5 (c 1.52 in CH₂Cl₂); ν_{max} (film)/cm⁻¹ 3432, 3239, 3042, 2953, 1736, 1712, 1627, 1540, 1455, 1417, 1439, 1374, 1282, 1256,

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1216, 1194, 1171, 1156, 1136, 1100, 1081, 1042, 1020, 107, 953, 917, 876, 756 and 701; δ_H (400 MHz, $CDCl_3$) 1.33-1.53 (3H, br m, $Pro\alpha-CH_3$), 1.62-2.20 (11H, m, $Pro\beta-H_2$, $Pro\gamma-H_2$ and $7\times$ cyclopentyl-H), 2.59-2.71 (1H, br m, $1\times$ cyclopentyl-H), 3.31-3.42 (1H, br m, $Pro\delta-H_AH_B$), 3.58-3.79 (4H, br m, OCH_3 and $Pro\delta-H_AH_B$), 4.92-5.17 (3H, m, $N-H$ and OCH_2Ph) and 7.27-7.42 (5H, s, Ph); δ_C (100 MHz, $CDCl_3$) 21.7 (CH_3 , $Pro\alpha-CH_3$), 24.1* (CH_2 , cyclopentyl-C), 24.2 (CH_2 , cyclopentyl-C), 24.4 (CH_2 , $Pro\gamma-C$), 24.5 (CH_2 , cyclopentyl-C), 36.4 (CH_2 , cyclopentyl-C), 37.1 (CH_2 , cyclopentyl-C), 37.2* (CH_2 , cyclopentyl-C), 37.7 (CH_2 , $Pro\beta-C$), 38.2* (CH_2 , cyclopentyl-C), 48.5 (CH_2 , $Pro\delta-C$), 52.1 (CH_3 , OCH_3), 66.6 (CH_2 , OCH_2Ph), 66.9 (quat., $Pro\alpha-C$), 67.2 (quat., $Gly\alpha-C$), 127.8 (CH, Ph), 128.2 (CH, Ph), 128.4 (CH, Ph), 136.6 (quat., Ph), 154.1 (quat., NCO_2), 155.7* (quat., NCO_2), 170.5 (quat., $Gly-CO$) and 174.7 (quat., CO_2CH_3); m/z ($El+$) 388.1991 (M^+ , $C_{21}H_{28}N_2O_5$ requires 388.1998).

(8a*S*)-Methyl-spiro[cyclopentane-1,3(4*H*)-tetrahydropyrrolo[7,2-*a*]pyrazine]-1,4(2*H*)-dione (Cyclic cyclopentyl-G-2-MeP)

To a solution of amide 22 (54 mg, 0.14 mmol) in methanol (4.6 cm^3) was added 10% Pd on activated charcoal (2.2 mg, 0.021 mmol) and the vessel flushed with hydrogen gas. The resulting suspension was stirred vigorously under an atmosphere of hydrogen for 17 h, then filtered through a CeliteTM pad with methanol (15 cm^3). The filtrate was concentrated to dryness under reduced pressure to give a yellow semi-solid which was purified by reverse-phase C18 flash column chromatography (0-10% CH_3CN/H_2O ; gradient elution) to produce cyclic cyclopentyl-G-2MeP (20 mg, 65%) as a yellow solid: mp 160-163° C.; $[\alpha]_D -97.9$ (c 1.61 in CH_2Cl_2); ν_{max} (film)/ cm^{-1} 3429, 2956, 2928, 2856, 1667, 1643, 1463, 1432, 1373, 1339, 1254, 1224, 1175, 1086, 1048, 976, 835, 774 and 730; δ_H (300 MHz, $CDCl_3$) 1.47 (3H, br s, $8a-CH_3$), 1.56-2.19 (11H, m, $8-H_2$, $7-H_2$ and $7\times$ cyclopentyl), 2.58-2.67 (1H, br m, $1\times$ cyclopentyl), 3.48-3.56 (1H, m, $6-H_AH_B$), 3.72-3.82 (1H, m, $6-H_AH_B$) and 6.56 (1H, br s, $N-H$); δ_C (75 MHz, $CDCl_3$) 19.9 (CH_2 , 7-C), 24.6 (CH_2 , cyclopentyl), 24.92 (CH_3 , $8a-CH_3$), 24.93 (CH_2 , cyclopentyl), 36.0 (CH_2 , 8-C), 38.7 (CH_2 , cyclopentyl), 41.9 (CH_2 , cyclopentyl), 44.8 (CH_2 , 6-C), 64.3 (quat., $8a-C$), 66.8 (quat., 3-C), 168.3 (quat., 4-C) and 172.2 (quat., 1-C); m/z ($El+$) 222.1369 (M^+ , $C_{12}H_{18}N_2O_2$ requires 222.1368).

In Vivo Testing

The following pharmacological studies demonstrate efficacy of cyclic G-2AllylP in attenuation of cognitive impairment. They are not intended to be limiting, and other compositions and methods of this invention can be developed without undue experimentation. All of those compositions and methods are considered to be part of this invention. All the following experiments were carried out using protocols developed under guidelines approved by the University of Auckland Animal Ethics Committee.

Efficacy of nootropic drugs can be conveniently tested using models of cholinergic hypofunction. Cholinergic hypofunction has been shown to contribute to dementia-related cognitive decline and remains a target of therapeutic intervention for Alzheimer's disease (Hunter 2004). The cholinergic hypofunction model is also applicable to other conditions. For example, it has been shown that scopolamine-induced cholinergic hypofunction can selectively impair the recognition accuracy of disgust and anger facial expressions rendering the of scopolamine on emotion-recognition similar to those found in Huntington's disease patients (Kamboy 2006). Scopolamine has been commonly used to induce cho-

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linergic hypofunction, and is a well-known model for human Alzheimer's disease, aging and other disorders of cognitive function (Liskowsky et al, Int. J. Dev. Neurosci, 24(2-3):149-156 (2006), Lindner et al., Psychopharmacology (Berl.) September 27 (2006), Bouger et al., Eur. Neuropsychopharmacol 15(3):331-346 (2005), Ebert et al, Eur. J. Clin. Invest., 28(11): 944-949 (1998), Barker et al, Int. J. Geriatr. Psychiatry, 13(4): 244-247 (1998), G. Smith, Brain Res. 471(2):103-118 (1998), Flood et al, Behav. Neural. Biol. 45(2):169-184 (1986)).

Example 5

Morris Water Maze (MWM) Model of Learning and Memory Used to Assess Effects of Cyclic G-2-AllylP on Cognitive Function

The purpose of the study was to investigate cyclic G-2AllylP in modes of cognitive deficit and affective state (anxiety).

20 Methods

The first part of the study involved acute testing of the cG-2AllylP in the Morris Water Maze memory model. The MWM test is one of the most frequently used tests for assessing spatial memory in rats and is well recognized to accurately predict effects of disease and treatment on spatial memory generally. Therefore, the MWM test reflects effects of disease and treatment in human subjects.

The standard procedure for MWM was followed. We used a circular swimming pool (80 cm depth \times 150 cm diameter) filled with opaque water, with the temperature maintained at 20° C. A platform was hidden 1 cm below the water surface, with a white flag (10 cm \times 10 cm) located either 20 cm above the platform for the visual cue and at 3 o'clock position in relation to the starting location for a spatial cue. On days 1-4 of the experiment rats underwent memory acquisition trials with 6 trials (60 seconds each) in each day of testing (habituation phase). Latency to reach the platform was recorded and the daily reduction of average latency was used to measure the capability to learn where the hidden platform was.

On day 5 of the experiment normal, non-aged Wistar rats were split into groups to receive either saline (n=28) or scopolamine (0.5 mg/kg, i.p., n=27) to induce memory deficit. Scopolamine was administered half an hour before the probe test commenced.

10 min following the scopolamine treatment, the cyclic G-2AllylP was administered orally at 30 mg/kg (n=31) with vehicle-treated animals administered the diluent by oral gavage using an identical treatment protocol (n=24).

Acute effects of cG-2AllylP were then tested in animals with scopolamine-induced memory impairment and in age-matched control animals with no memory impairment to determine any direct pharmacological effect on memory processing. Experimental groups are detailed in the Table 1 below.

TABLE 1

Animals Used to Test Effects of cG-2-AllylP on Memory		
	Scopolamine	Vehicle
Vehicle	N = 12	N = 12
cG-2AllylP	N = 15	N = 16

On day 5, the probe MWM test was performed with the platform removed. There were 6 trials, each of maximum duration of 60 s, at least 5 min rest between trials). The amount of time the rats spend swimming near the platform

provided a measure of how much they relied on visual and spatial cue to locate the platform, as opposed to using a non-spatial strategy. Data was collected and analysed using Any-maze (v4.2) software.

The data generated from behavioural tests was analysed using one-way ANOVA for determining the difference between the aged-groups. Two-way ANOVA was used for examining the progress of behavioral results with the time points treated as dependent factors. GraphPad Prism version 3.02 was used for data analysis.

Results

Treatment with scopolamine significantly impaired acquisition of spatial memory in treated animals (time to platform approximately 208% of control on day 4). Cyclic G-2-AllylP (30 mg/kg; daily) significantly reversed the cognitive impairment induced by scopolamine (FIGS. 1A, 1B, 1C).

Example 6

cG-2-AllylP Improves Synaptic Plasticity and Aging-Related Memory Loss Methods

Aged rats (male Wistar rats, 18-20 months old) were divided into four groups: two vehicle-treated (groups 1 and 3) and two G-2-AllylP treated (groups 2 and 4) (all groups n=6-8). Cyclic G-2-AllylP was synthesised by the Department of Medicinal Chemistry and dissolved in normal saline before the treatment. On day 1 a single dose of cyclic G-2-AllylP was given centrally (20 ng/animal, i.c.v.) to the animals in groups 2 and 4; saline was administered to groups 1 and 3. The memory tests using Novel Object Recognition Test started either on day 3 (groups 1 and 2) or 24 (groups 3 and 4) after the treatment. On the completion of the NORT, the rats were killed with an overdose of sodium pentobarbital and were perfused transcardially with normal saline followed by 10% formalin. Tissues collected at day 7 in groups 1 and 2, and at day 28 from groups 3 and 4. The brains were kept in the same fixative for a minimum of 2 days before being processed using a standard paraffin embedding procedure. Briefly, small blocks (10x10x3 mm) of tissue were fixed for up to 24 hrs. The blocks were then infiltrated and embedded with paraffin and cut in ribbons and mounted on slides. Slides were then stored until immunostaining was commenced. Synaptogenesis in brain tissue was examined using immunohistochemical staining.

Novel Object Recognition Test (NORT)

Exploratory activity is a typical learning behaviour displayed by animals including humans and rats in novel environments. Exploratory activity decreases over time when the novel becomes familiar and the habituation occurs. In familiar environments, exploratory activity can be reactivated by introducing a novel object. The increase in exploring behaviour once the environment is altered following a habituation provides a measure of the memory for the familiarity and the recognition of the novelty.

In this Example, we carried out two NORTs, one at days 3-6 and the other at 24-27 days. The rats were allowed to familiarise themselves with the testing arena (90x60x40 cm) in the first day of NORT. In the following two days of each test, four novel objects were placed into the testing arena and the rats had 2 trials each day (each of 15 min duration and 2 hours apart). The time spent on exploring the objects was reduced once the animal tested learned about the objects (training phase). In the last day (day 4 of each test), one familiar object was replaced by a novel object before the second trial (test 6, testing phase). The average time spent on exploring the 3 familiar objects and the time spent on explo-

ration of the novel object was used as a measure for the memory of familiarity and the novelty recognition.

Effects of cG-2-AllylP on Expression of NMDA Receptors, AMPA Receptors, rKrox-24 and Synaptophysin mRNA in the Hippocampus

It is accepted that the hippocampal formations in humans and animals play a crucial role in a number of memory types (Morris et al. 2006 *Europ. J. Neurosci.* 23, 2829). The specific functionality remains under dispute, but there is an understanding that hippocampus plays a key role in the automatic encoding and initial storage of attended experiences (episodic memory formation), memory consolidation and novelty detection. The first aspect, encoding and short-term storage of memories, is dependant on the synaptic plasticity and synaptic transmission, both of which are linked to glutaminergic neurotransmission.

Glutaminergic transmission is facilitated by two types of glutamate receptors: N-methyl-D-aspartate receptors (NMDAR) and a-amino-3-hydroxy-5-methyl-4-isoxalone propionic acid receptors AMPA or non-NMDA receptors.

APMA receptor subunit GluR1 is a post-synaptic receptor and has been commonly used for memory measurement. GluR1 is believed to mediate calcium influx, and has a vital function in synaptic plasticity related to learning. It has been previously suggested (Hayashi et al., 2000) that incorporation of GluR1 into synapses might be important for long-term potentiation (LTP), which is essential for learning and memory.

It had been demonstrated that NMDA receptor subunit NR1 is crucial for formation of spatial memory. In knock-out models where the R1 subunit of the NMDA receptor in the pyramidal cells of the CA1 region was selectively knocked-out, the long-term potentiation was shown to be abolished (Tsien 1996).

Synaptophysin is a presynaptic vesicle protein. Its quantitative detection is established as a molecular marker of synaptic density.

The neuronal transcript factor Krox24 staining is used as a marker for neuronal plasticity. The protein products of the Krox24 family (as well as by brain-derived neurotrophic factor, BDNF) have recently linked with stabilizing synaptic modifications occurring during NMDA-receptor-mediated hippocampal LTP and LTD. (Dragunow. 2006. *Behaviour genetics.* 23; 293).

Immunohistochemical Staining

Conventional deparaffinisation and rehydration techniques were used to allow the water-based buffers and antibodies to penetrate the tissue slices. Antigen retrieval was used only prior to AMPA receptor (GluR1) staining, i.e. the slides were placed in boiling citrate buffer and allowed to cool.

The following antibodies were used:

- i) primary rabbit antibody to NMDA NR1 subunit, at 1:200 concentration in buffer, incubated for 48 hrs (Chemicon—AB1516), followed by Sigma fluorescent secondary antibody (alexaFluor 594), at 1:200 dilution, incubated for 24 hours at 40 C.
- ii) primary antibody to AMPA GluR1 subunit, at 1:50 concentration in buffer, incubated for 48 hrs (Chemicon—AB1504) followed by 3,3'-diaminobenzidine (DAB) at 1:200 dilution, incubated for 24 hours at 40 C.
- iii) Primary antibody to mSynaptophysin (Sigma—S5768), at 1:200 concentration in buffer, followed by DAB at 1:200 dilution, incubated for 24 hours at 40 C).
- iv) Primary antibody to rKrox-24 (Santa Cruz—catalogue number SC-189) at 1:200 concentration in the buffer,

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followed by anti-rabbit secondary antibody at concentration of 1:200 dilution, incubated for 24 hours at 40 C. (e) Antibodies were detected using light microscopy.

Results NORT

A trend to improve the novelty recognition in the groups treated with cG-2-AllylP was observed after 27 days (FIG. 2), but not 6 days after the treatment (no figure). We conclude that the cG-2-AllylP treatment improved novelty recognition in the drug-treated animals at 27 days.

AMPA Glutamate Receptor-1 Staining

Hippocampal slices from regions CA1 (granular cell layer, strata oriens and radiatum) and CA3 (pyramidal cell layer) were stained for AMPA receptors GluR1.

In CA3 there was no change in the number of receptors in each region on either day 7 or 28. There was however a significant increase in the number of AMPA receptors in CA1 (granular cell layer) (FIG. 4) and CA1 stratum oriens (FIG. 5) and on day 28.

That histological change was correlated with the improved performance in the novel object recognition test. The improved memory (FIG. 2) was correlated to the elevated AMPA glutamate receptor-1 (FIG. 3). We concluded that cG-2-AllylP improved glutaminergic neuro-transmission (GluR1) at post-synaptic level.

We observed that cG-2-AllylP treatment resulted in a long term increase in GluR1 staining on the post-synapses and increased the density of pre-synaptic vesicles. As the majority of vesicles in the hippocampus are glutamic vesicles, we concluded that the long term memory improvement was associated with increased glutamic neurotransmission.

Synaptophysin Staining

We subsequently analysed effect of cG-2-AllylP on the levels of synaptophysin staining in CA3 and CA1 regions of the hippocampus.

In all tested areas there was either a significant increase (CA3) or a clear trend towards (CA1—strata oriens and radiatum) the increase in the density of synaptophysin staining at 28 days post-treatment. That increase is a marker of increased synaptic plasticity and a clear indication of synaptogenesis which is a most likely cause of the improvement in the performance of the treated groups in applied memory tests.

NMDA Receptor-1 Staining

While there is a significant improvement in AMPA receptors post-treatment, the changes in the NMDA receptors are not so pronounced (FIGS. 9A, 9B and 9C).

Krox24 Staining

We analysed the density of the Krox24 staining in the CA1-2 regions of the hippocampus. We observed a trend towards the increased density in treatment group in comparison to the vehicle treated group. We conclude that the Krox24 staining results positively correlate with improved memory function (FIG. 10).

Example 7

cG-2-AllylP Increases the Number in Pre-Synaptic Vesicles in the Hippocampus of Middle Aged Rats

Four middle aged Wistar male rats (12 months) were divided into two groups: one vehicle-treated (n=2) and one cG-2-AllylP-treated (n=2). The rats were treated subcutaneously with 3 mg/kg/day of either saline or cG-2-AllylP for 7 days. On day 21 of the experiment the animals were sacrificed and the hippocampal tissue was harvested. Semi-thin sections of the tissue were fixed with OsO₄ and embedded in resin. CA1 stratum oriens and CA3 sections were then sliced into

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ultra-thin, 80 nm slices and stained with uranyl acetate and lead citrate. Approximately 50 synapses per animal were analysed, synapse type classified and vesicle density was measured using AnalySIS®.

5 Transmission electron microscopy was used to count the total number of vesicles on the slides. The average density was calculated by measuring the total area (using AxioVision software) and using the number of vesicles. We followed the protocol in Yoshida et al. 97, Journal of Neurochemistry to calculate the vesicle density in a 200 nm×200 nm square apposing the post-synaptic density (PSD).

Results

The number of pre-synaptic vesicles in the CA1 and C3 subregions of the hippocampus was increased after cG-2-AllylP treatment (3 mg/kg/day×7 days, s.c.) compared to the vehicle treated animals at 21 days after the treatment (FIG. 11).

We conclude from these studies that scopolamine treatment can decrease cognitive function in animals, and that these changes can mimic cognitive impairment in human beings with one or more of a variety of neurological conditions. Additionally, we conclude that cG-2-AllylP can improve cognitive function in scopolamine-treated animals and in animals with normal aging-related cognitive impairment. Further, we conclude that cG-2-AllylP can increase synaptogenesis, increase AMPA receptors, increase neural plasticity, can stabilize synaptic modifications and can increase novelty recognition.

These studies therefore support the use of cG-2-AllylP as an effective pharmacological agent to treat a variety of cognitive impairments in animals including humans suffering from Alzheimer's disease, Parkinson's disease, and other chronic neural disorders, as well as cognitive impairment associated with aging.

35 In Vitro and In Vivo Testing

The following pharmacological studies demonstrate neuroprotective features of this invention. They are not intended to be limiting, and other compositions and methods of this invention can be developed without undue experimentation. All of those compositions and methods are considered to be part of this invention. All the following experiments were carried out using protocols developed under guidelines approved by the University of Auckland Animal Ethics Committee.

Example 8

Effects of Cyclic G-2-AllylP and Cyclic cyclopentyl-G-2MeP on Cerebellar Cell Explants

To determine the effects of cG-2-AllylP and cyclic cyclopentyl-G-2-MeP on neuronal cells in vitro, a series of studies was carried out using cerebellar explants from adult rats. In vitro systems are suitable for studying neuronal proliferation, neurite growth, formation of nerve bundles, and effects of toxins on neural cells, effects that parallel effects observed in vivo. Thus, results of studies using in vitro cerebellar explants are predictive of effects of interventions in vivo.

In a first series of studies, effects of glutamate on cerebellar explants were determined. At physiological concentrations, glutamate is a neurotransmitter in the CNS of mammals, including humans. However, at sufficiently high concentrations, glutamate is neurotoxic, resulting in neuronal cell death. Because glutamate is a naturally occurring neurotransmitter in the CNS of mammals, including humans, and because glutamate neurotoxicity is recognized in the art as reflective of neurotoxicity in general, and including cell death

and degeneration, it is a valuable tool useful for identifying and characterizing agents effective in treatment of neurodegeneration and neural cell death.

Materials and Methods

Cover slips were placed into a large Petri dish and washed in 70% alcohol for 5 minutes, then washed with Millipore H₂O. The cover slips were air dried, and coated with Poly-D-Lysine (1 mg/ml stock solution in PBS, 90-100 μ l) for 2 hours at 34° C.

Extraction of Cerebellar Tissue

Postnatal day 8 Wistar rats were used for the study. The rats were sacrificed and placed in ice for 1 minute, decapitated and the cerebellum removed and placed on ice. Cerebellum tissue was placed in 1 ml of 0.65% glucose-supplemented PBS (10 μ l 65% stock D (+)glucose/1 ml PBS) in a large Petri dish, chopped up into smaller sections and triturated with a 1 ml insulin syringe via a 23 G (0.4 mm) needle, and then squirted back into the glucose solution in the large Petri dish. The tissue was sieved through (125 μ m pore size gauze) and centrifuged (2 minutes at 60 g) twice to exchange the medium into serum-free BSA-supplemented START V medium (Biochrom, Germany). The second centrifugation step was done with 1 ml of START V medium. The microexplants were reconstituted into 500 μ l of START V medium and put on ice. Cultivation of Cerebellar Cells

Two hours after PDL-coating, the slides were washed with Millipore H₂O and air dried. Each slide was placed into a small Petri dish (diameter: 35 mm) and 40 μ l of START V/cell suspension was added. The tissue was incubated for 2 hours at 34° C. (settlement period). START V-medium (1 ml) was then added to the Petri dish and cultivated at 34° C. in the presence of 5% CO₂ in air at 100% humidity for 48 hours.

Drug Application

For the study, certain explant cultures were exposed to vehicle (PBS) only. In the first study (Study 1) 10 μ l of toxin 1 (L-glutamate-100 mM in Millipore water; final concentration: 1 mM) and 10 μ l of toxin 2 (3-nitropropionic acid—50 mM—pH 7—in Millipore water, final concentration: 0.5 mM) was applied simultaneously with the drug to be tested (10 mM stock solution prepared in PBS and diluted to final concentrations between 1-100 nM). In each case, the drugs were left in contact with the explants for the duration of the study.

Methods for Determining Drug Effects

After explants were exposed to drugs for the study period, cells were then rinsed in PBS and then fixed in increasing concentrations of paraformaldehyde (500 μ l of 0.4% PFA was applied; then 1.2% PFA; then 3% PFA and finally 4% PFA (each fixation step: 2-3 minutes). Finally, the microexplants were rinsed in PBS.

Neurons in the explants were then evaluated for morphology (presence of neurites) and counted as live cells per microscopic field. Four fields displaying highest cell density were counted per cover slip and the data presented as mean \pm standard error of the mean (SEM); n=4 each. Statistical significance was evaluated by using the non-paired Student's t-test.

Results

Cyclic G-2-AllylP

The results of the study are shown in FIG. 12. Glutamate treatment (1 mM; filled bar) resulted in about an 85% loss of cerebellar neurons having neurites compared to vehicle-treated controls (open bar). In contrast, cG-2AllylP significantly increased the numbers of cells having neurites in a dose-dependent manner when administered simultaneously with glutamate (shaded bars). Treatment with low doses of

cG-2AllylP (100 pm to 10 nm) showed a significant decrease in glutamate-induced neurotoxicity.

Cyclic cyclopentyl-G-2-MeP

The results of the study are shown in FIG. 13. Cyclic cyclopentyl-G-2MeP significantly increased the number of cells having neurites when simultaneously administered with glutamate (light shaded bars). Treatment with low doses of cyclic cyclopentyl-G-2MeP showed a significant decrease in glutamate-induced neurotoxicity.

Conclusions

Both cG-2-AllylP and cyclic cyclopentyl-G-2-MeP independently decreased or prevented glutamate-induced neurotoxicity, indicating that both drugs are neuroprotective and can be used to inhibit neuronal degeneration or cell death.

Example 9

Effects of cG-2-AllylP on Hypoxic-Ischemic Injury I

Materials and Methods

To determine whether cG-2AllylP might prevent neuronal injury in response to stroke, cardiac arterial bypass graft surgery (CABG) or other hypoxic insults, a series of studies were carried out in rats that had been exposed to hypoxic-ischemic injury (HI). Adult rats (Wistar, 280-310 g, male) were used. The modified Levine model preparation and experimental procedures were used (Rice et al, 1981, *Ann. Neurol.*: 9: 131-141; Guan et al *J.*, 1993, *Cereb. Blood Flow Metab.*: 13(4): 609-16). These procedures in brief, consist of an HI injury induced by unilateral carotid artery ligation followed by inhalational asphyxia in the animals with an implanted lateral ventricular cannula. A guide cannula was stereotactically placed on the top of the dura 1.5 mm to the right of the mid-line and 7.5 mm anterior to the interaural zero plane under halothane anaesthesia. The right carotid artery was double ligated two days after the cannulation. After 1 hour recovery from the anaesthesia, each of the rats were placed in an incubator where the humidity (90 \pm 5%) and temperature (31 \pm 0.5° C.) were controlled for another hour, then exposed to hypoxia (6% oxygen) for 10 min. The animals were kept in the incubator for an additional 2 hours before treatment.

Nine pairs of rats were treated intracerebrally (icv) with either cG-2AllylP (2 ng) or its vehicle (normal saline) 2 hours after hypoxic-ischemic insult. Rats in each group were simultaneously infused with cG-2-AllylP or its vehicle under light anaesthesia (1.5% halothane) 2 hours after the insult. A total volume of 20 μ l was infused (icv) over 20 minutes by a micro-infusion pump.

Histological examination was performed on rats 5 days after the hypoxic-ischemic injury. The rats were killed with an overdose of sodium pentobarbital and were perfused transcardially with normal saline followed by 10% formalin. The brains were kept in the same fixative for a minimum of 2 days before being processed using a standard paraffin imbedding procedure.

Coronal sections 8 μ m in thickness were cut from the striatum, cerebral cortex and hippocampus and were stained with thionin and acid fuchsin. The histological outcome was assessed at three levels: (1) the mid level of the striatum, (2) where the completed hippocampus first appeared and (3) the level where the ventral horn of the hippocampus just appears. The severity of tissue damage was scored in the striatum, cortex and the CA1-2, CA3, CA4 and dentate gyrus of the hippocampus. Tissue damage was identified as neuronal loss

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(acidophilic (red) cytoplasm and contracted nuclei), pan-necrosis and cellular reactions. Tissue damage was scored using the following scoring system: 0: tissue showed no tissue damage, 1: <5% tissue was damaged, 2: <50% tissue was damaged, 3: >50% tissue was damaged and 4: >95% tissue was damaged.

Results and Conclusion

The results of this study are shown in FIG. 14. FIG. 14 shows that hypoxic-ischemic injury (left bars of each set) resulted in significant damage scores in each of the areas of the brain studied. FIG. 14 also shows that central administration of a relatively low dose of cG-2-AllylP (right bars of each set; 2 ng) significantly reduced the tissue damage in each brain region examined compared to the vehicle treated group ($p < 0.001$).

It can be seen that cG-2-AllylP can be neuroprotective against neural damage caused by hypoxic-ischemic injury, even when administered after hypoxic-ischemic injury. This surprising finding indicates that cG-2-AllylP is a useful agent to treat a variety of conditions characterized by neural degeneration or cell death.

Example 10

Effects of cG-2-AllylP on Hypoxic-Ischemic Injury II

Materials and Methods

Materials and methods described in Example 9 were used and the number of treatment groups was increased. Rats were divided into 5 treatment groups treated intracerebrally (icv) with one of 4 doses of cG-2-AllylP or with its vehicle (normal saline) 2 hours after hypoxic-ischemic insult (1: $n=10$, 2 ng; 2: $n=9$, 4 ng; 3: $n=9$, ng; 4: $n=10$, 100 ng; and 5: $n=9$, vehicle).

FIG. 15 shows hypoxia alone (vehicle) produces neuronal damage scores in all areas of the brain studied. In animals treated with cG-2-AllylP, hypoxia had less effect, even though the agent was administered after the hypoxic/ischemic injury. The neuroprotective effect was observed for all doses of cG-2-AllylP, except for the highest dose (100 ng) administered to the striatum. However, in all other sites and with all other doses, cG-2-AllylP lessened the neural damage effects of hypoxia/ischemia. Moreover, cG-2-AllylP had an increased efficacy in brain regions that experienced progressive injury associated with delayed cell death, such as that associated with apoptosis. In brain regions such as the dentate gyrus and the cerebral cortex, that are more resistant to HI injury, the progression of injury is known to be slower and more severe than in the brain regions that are more sensitive to HI injury such as the striatum and the CA1-2, CA3 and CA4 subregions of the hippocampus. This result shows that cG-2-AllylP can be beneficial in treatment of chronic neurological disorders.

This invention is described with reference to specific embodiments thereof. Other features and embodiments of this invention can be produced by those of skill in the art without undue experimentation and a reasonably likelihood of success. All of those embodiments are considered to be part of this invention.

What is claimed is:

1. A method for relieving or alleviating a symptom of cognitive dysfunction caused by a disease, injury, or condition in a mammal in need thereof, comprising: administering a pharmaceutically effective amount of cyclic Glycyl-2-Allyl Proline (cG-2-AllylP) to said mammal thereby providing

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relief from the symptom, where said symptom is loss of spatial memory, loss of long-term memory, or loss of novelty recognition.

2. The method of claim 1, where said disease is Alzheimer's disease.

3. The method of claim 1, where said disease is Huntington's disease.

4. The method of claim 1, where said disease is Lewy Body disease.

5. The method of claim 1, where said disease is Dementia.

6. The method of claim 1, where said disease is cerebral atrophy.

7. The method of claim 1, where said disease is frontotemporal lobar degeneration.

8. The method of claim 1, where said disease is Pick's disease.

9. The method of claim 1, where said disease is multi-infarct dementia.

10. The method of claim 1, where said disease is HIV infection.

11. The method of claim 1, where said disease is Down's syndrome.

12. The method of claim 1, where said injury is neurotoxic injury.

13. The method of claim 1, where said injury is cerebral hypoxia/ischemia.

14. The method of claim 1, where said injury is traumatic brain injury.

15. The method of claim 1, where said injury is coronary artery bypass surgery.

16. The method of claim 1, where said condition is normal aging.

17. The method of claim 1, where said condition is age-related memory loss.

18. The method of claim 1, where said condition is cholinergic hypofunction.

19. The method of claim 18, where said cholinergic hypofunction is caused by scopolamine.

20. The method of claim 1, wherein said cognitive dysfunction is caused by a decrease in glutamate receptors in the granular cell layer (CA1) of the hippocampus of said mammal.

21. The method of claim 1, where said condition is vascular narrowing or blockage in the brain.

22. The method of claim 1, said condition is decreased cerebral perfusion.

23. The method of claim 1, where said condition is, neuroinflammation.

24. The method of claim 1, where said condition is mild cognitive impairment.

25. The method of claim 1, where said condition is loss of synaptic plasticity.

26. The method of claim 1, wherein said cG-2-AllylP comprises an aqueous solution and one or more pharmaceutically acceptable excipients, additives, carriers or adjuvants.

27. The method of claim 1, wherein said cG-2-AllylP further comprises one or more excipients, carriers, additives, adjuvants, or binders in a tablet or capsule.

28. The method of claim 1, wherein said cyclic G-2-AllylP is administered via an oral, intraperitoneal, intravascular, peripheral circulation, subcutaneous, intraorbital, ophthalmic, intraspinal, intracisternal, topical, infusion, implant, aerosol, inhalation, scarification, intracapsular, intramuscular, intranasal, buccal, transdermal, pulmonary, rectal, or vaginal route.

29. The method of claim 1, wherein said effective amount has a lower limit of about 0.001 milligrams per kilogram mass (mg/kg) of the animal and an upper limit of about 100 mg/kg.

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